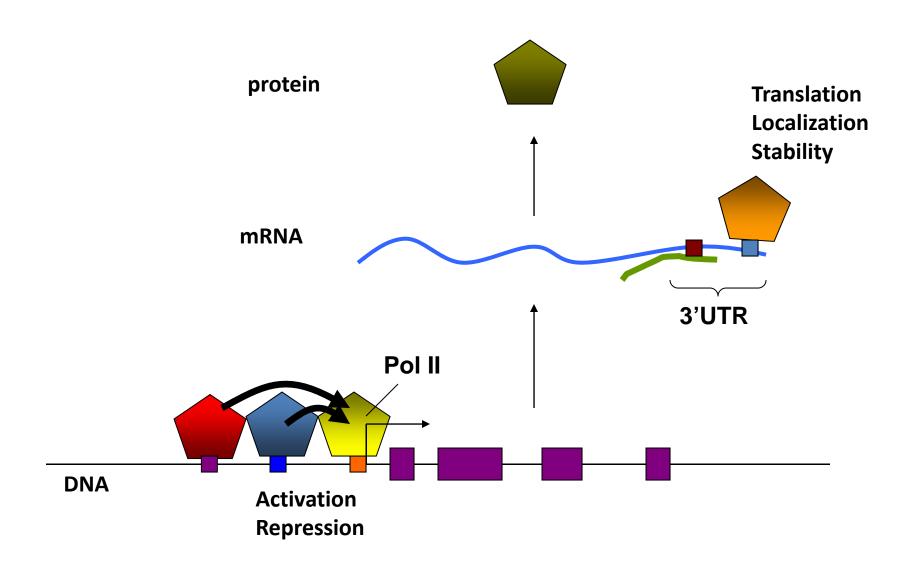
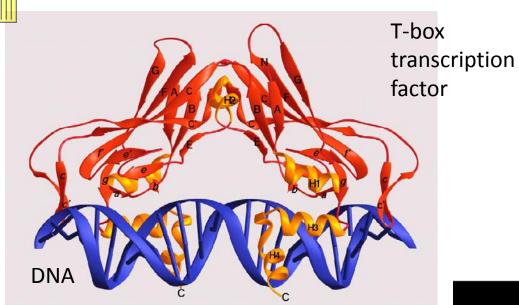
# Discovering regulatory sequences from expression data

- Unsupervised clustering
- Information theory
- Optimization
- Non-parametric statistical testing
- Multiple testing
- Overfitting

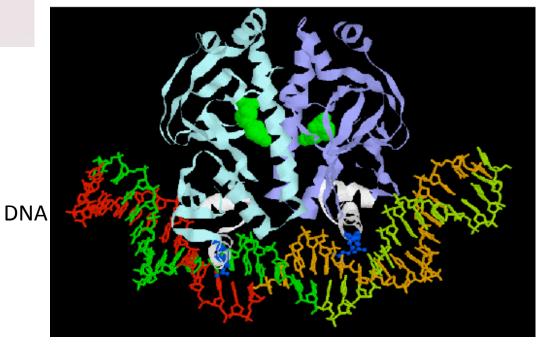


## Transcriptional and post-transcriptional regulation of gene expression

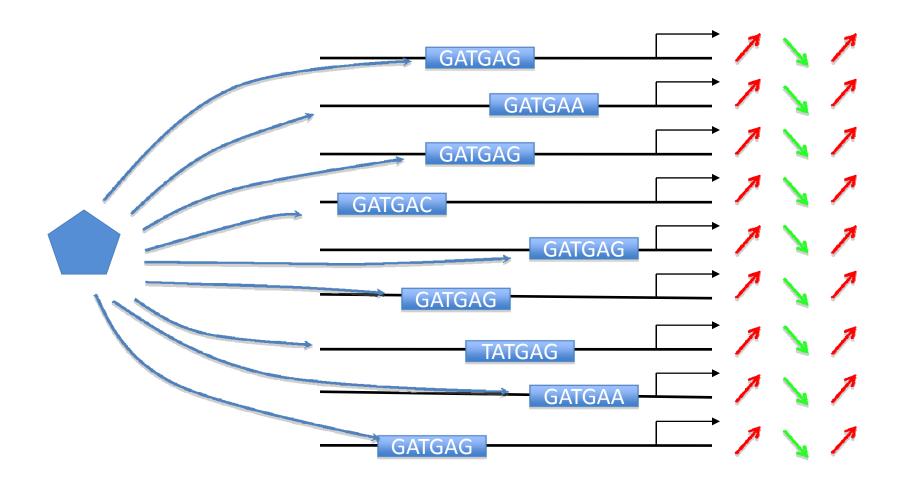




CRP

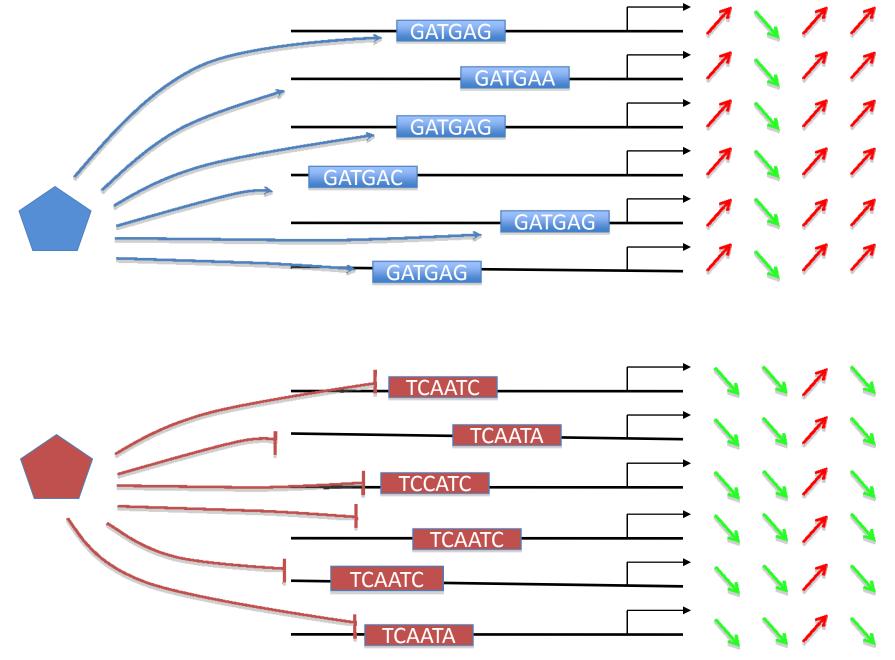


Transcription factor binding sites are ~6-12 bp-long

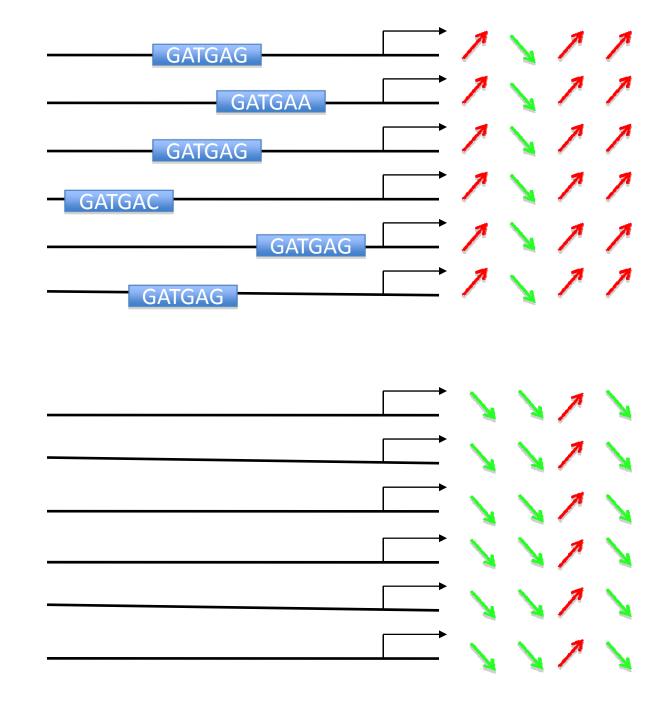


Genes regulated by the same TF will be co-expressed

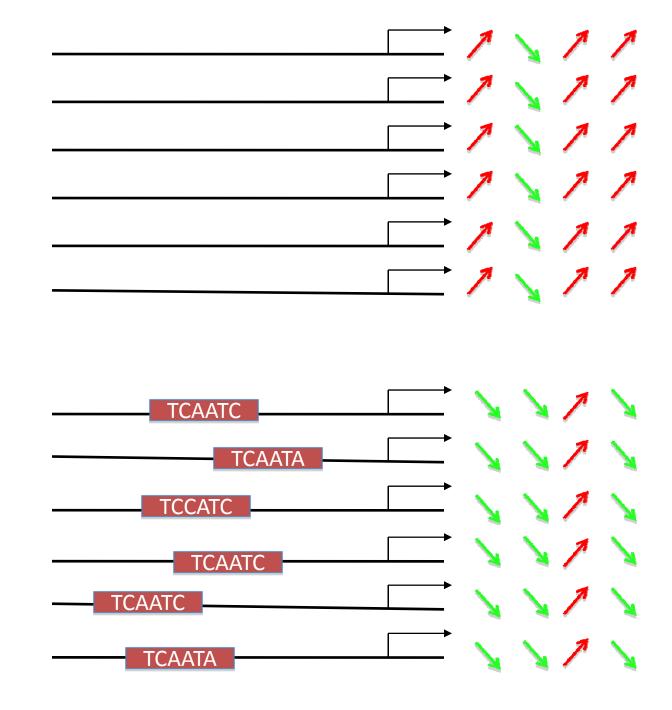








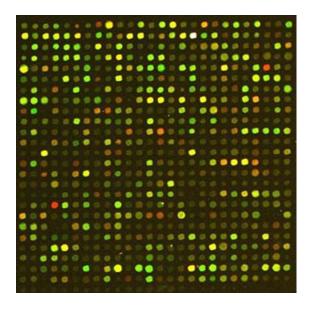


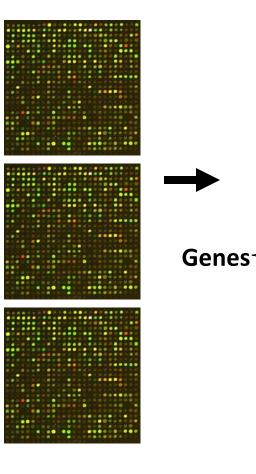


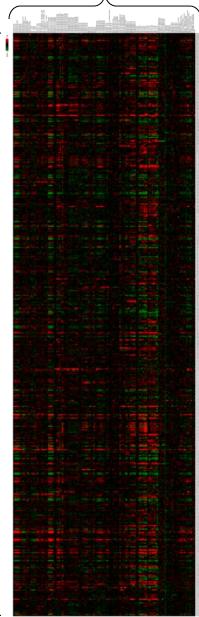
Microarray Experiments

Several microarray experiments (conditions, time points, treatments)

#### Microarray experiment







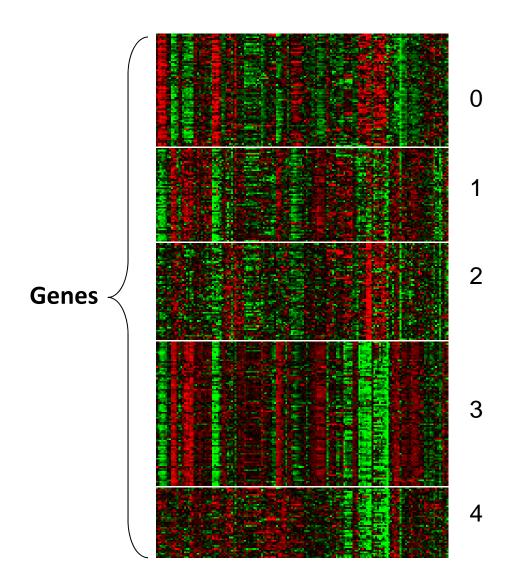
...

### Creating co-expression clusters

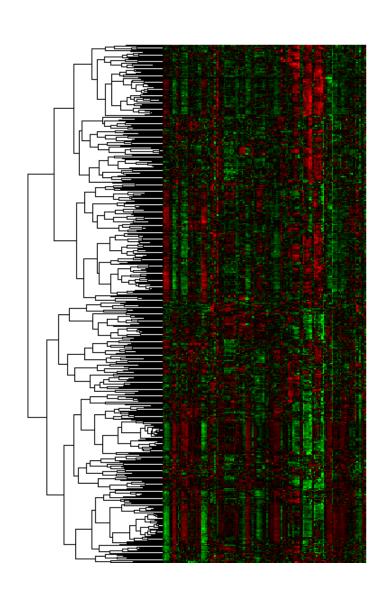
Unsupervised clustering approaches:

- K-means
- Self-organizing maps
- Hierarchical clustering

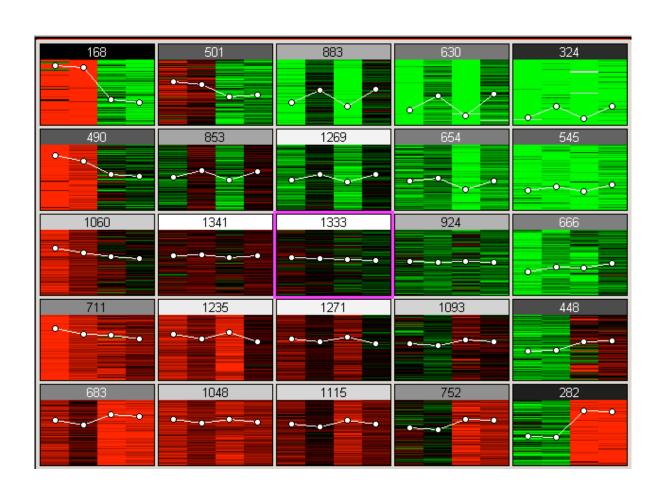
## K-means clustering



## Hierarchical clustering

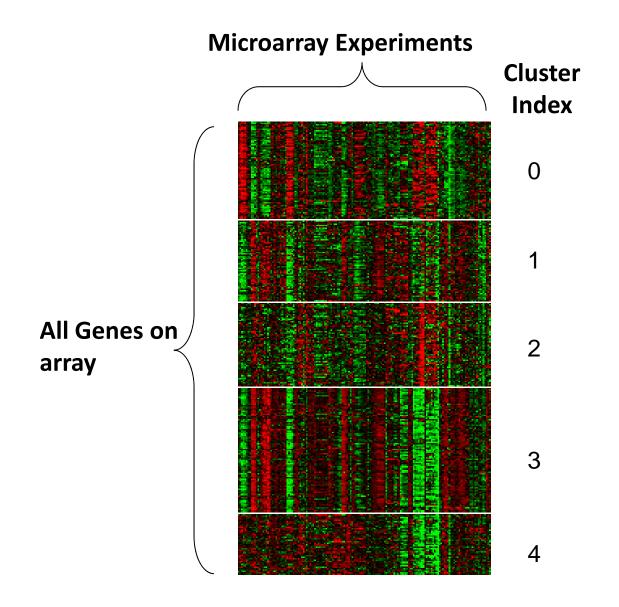


## Self-organizing map





## Clusters of co-expressed genes





#### A Universal Framework for Regulatory Element Discovery across All Genomes and Data Types

Olivier Elemento, 1,2,3 Noam Slonim, 1,2,3,4 and Saeed Tavazoie 1,2,\*

Princeton University, Princeton, NJ 08544, USA

DOI 10.1016/j.molcel.2007.09.027

#### SUMMARY

Deciphering the noncoding regulatory genome has proved a formidable challenge. Despite the wealth of available gene expression data, there currently exists no broadly applicable method for characterizing the regulatory elements that shape the rich underlying dynamics. We present a general framework for detecting such regulatory DNA and RNA motifs that relies on directly assessing the mutual information between sequence and gene expression mea-

specific short DNA sequences and then act to modulate the activity of the RNA polymerase. Transcript stability, localization, and translation are also regulated by proteins and RNAs (e.g., miRNAs), which also bind specific short RNA sequences, generally in 3'UTRs. A comprehensive characterization of these DNA and RNA regulatory elements is a formidable challenge, especially within complex metazoan genomes. Experimental (Gerber et al., 2004; Harbison et al., 2004) and computational approaches are emerging to meet these challenges. Several methods compare the intergenic regions of different genomes, aiming to detect sequence elements that are highly conserved across related species (Elemento and Tavazoie, 2005;

<sup>&</sup>lt;sup>1</sup>Lewis-Sigler Institute for Integrative Genomics

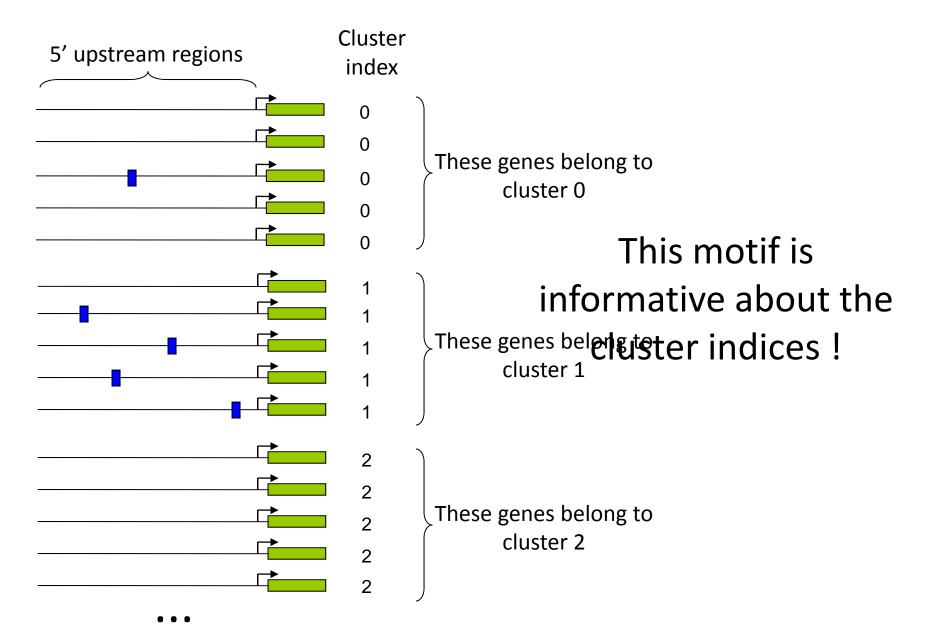
<sup>&</sup>lt;sup>2</sup>Department of Molecular Biology

<sup>&</sup>lt;sup>3</sup>These authors contributed equally to this work.

<sup>&</sup>lt;sup>4</sup>Present address: IBM Haifa Research Labs, Haifa 31905, Israel.

<sup>\*</sup>Correspondence: tavazoie@genomics.princeton.edu



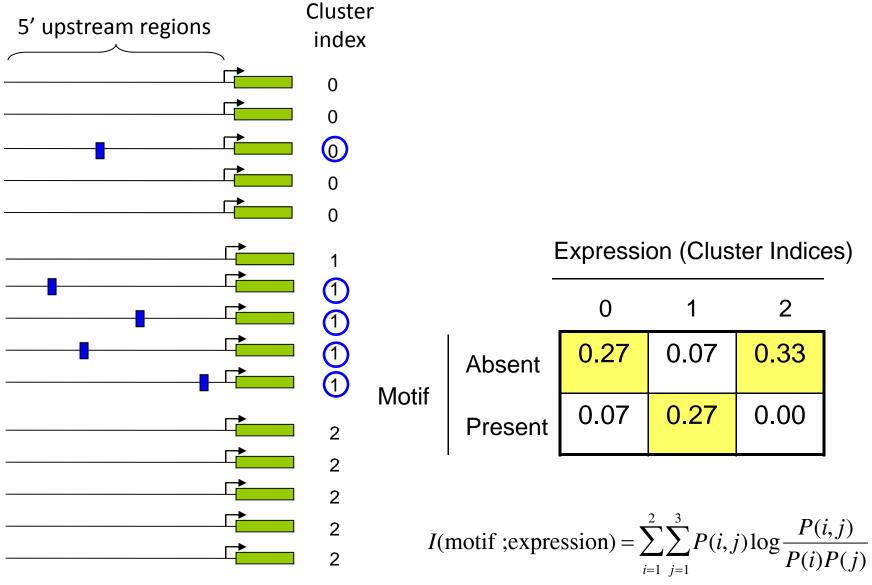


### Mutual Information

$$I(X;Y) = \sum_{x} \sum_{y} P(x,y) \log \frac{P(x,y)}{P(x)P(y)}$$

I(X;Y) quantifies the amount of information that a variable X contains about another variable Y

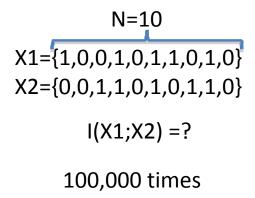


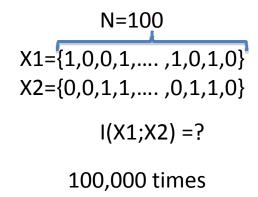


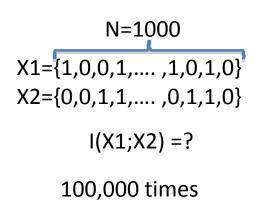
• • •

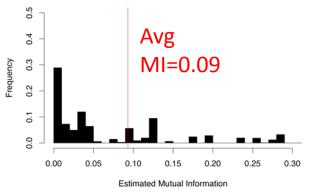
$$I(M;E) = 0.34 bits$$

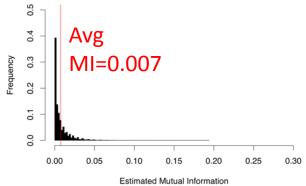
# MI estimator is biased (sample size bias)

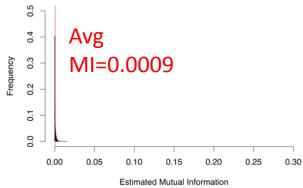












# MI estimator is biased (sample size bias)

Can correct for sample size bias, e.g. Slonim et al,
 2002 ... slow ... not very precise ... not necessary if:

 Keep sample size the same so that we can compare MI values

 Estimate how large an MI value is compared to expected MI



# Algorithm for finding informative motifs

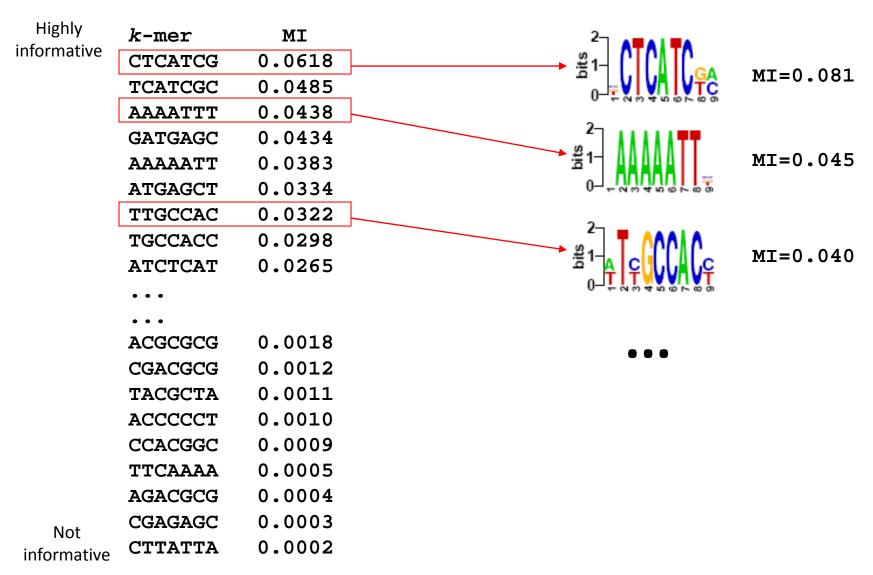


## motif representations

	Accuracy	Search space	
Degenerate code	good	large	
[AC]CGATGAG[TC]	god		
Words ( <i>k</i> -mers)	acceptable	small	
GCGATGAG	acceptable		

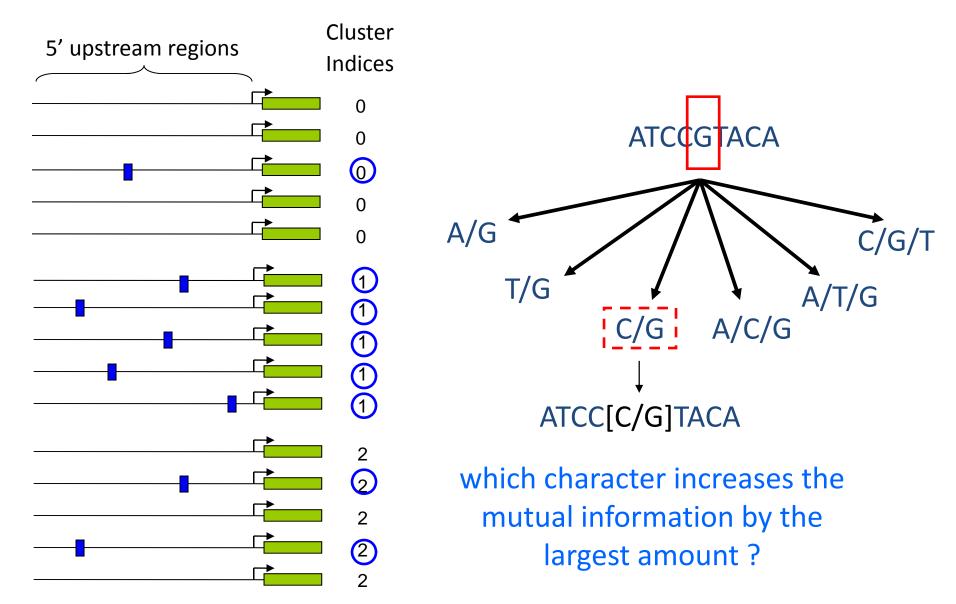


## Motif Search Algorithm



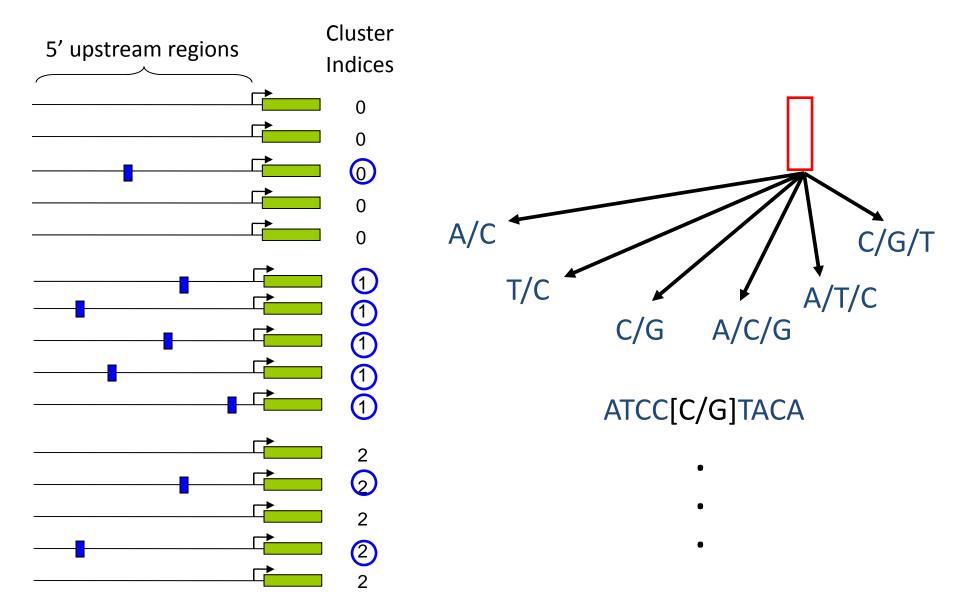
### 1

# Optimizing k-mers into more informative degenerate motifs



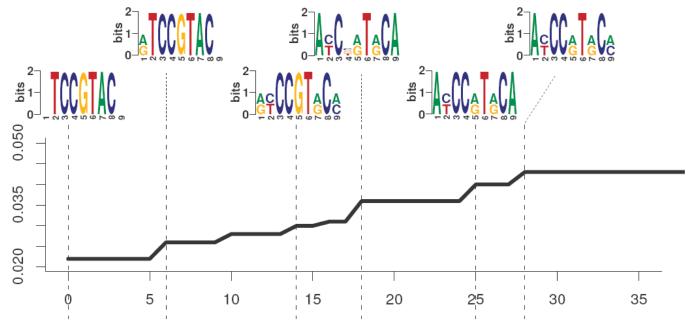


# Optimizing k-mers into more informative degenerate motifs



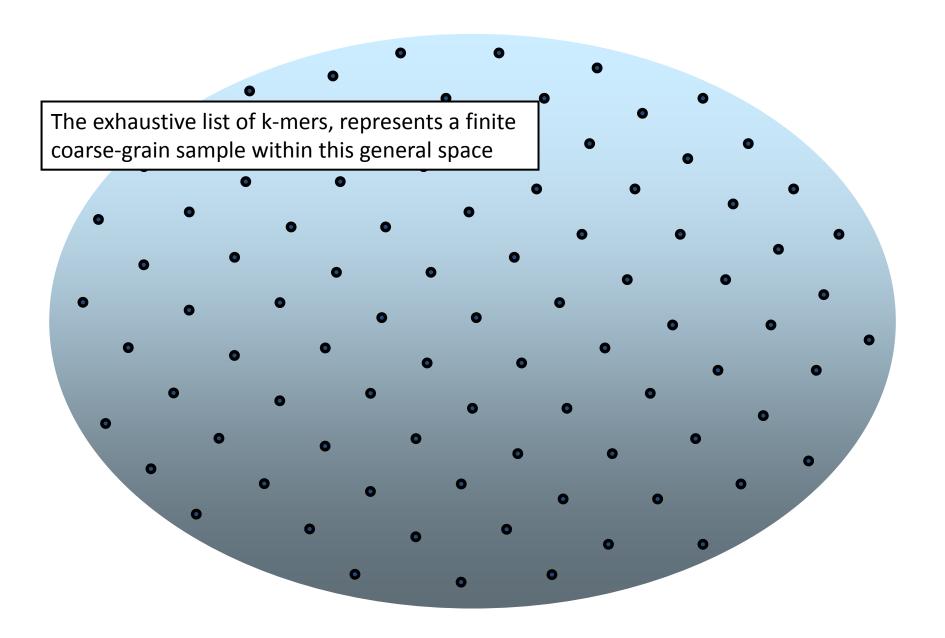
1



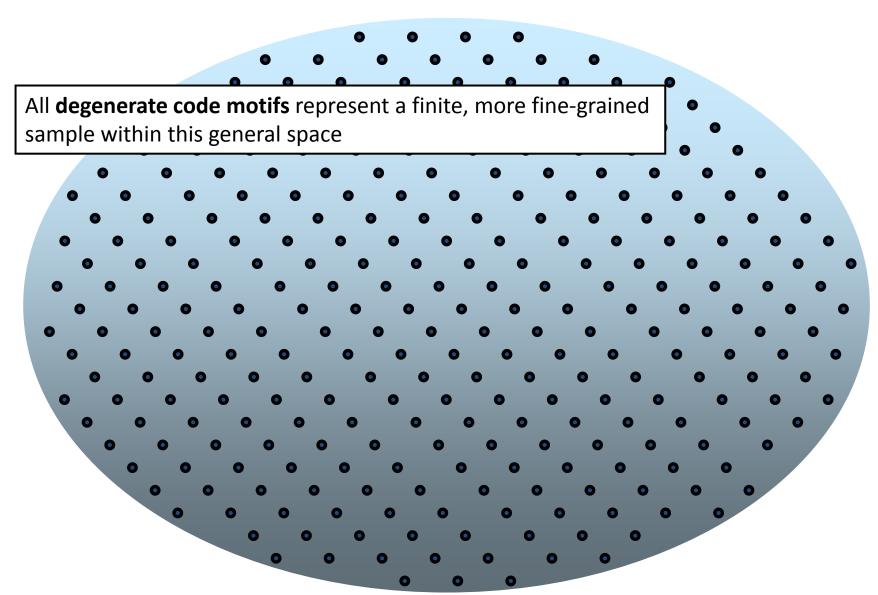


Optimization iterations (visited motif positions)

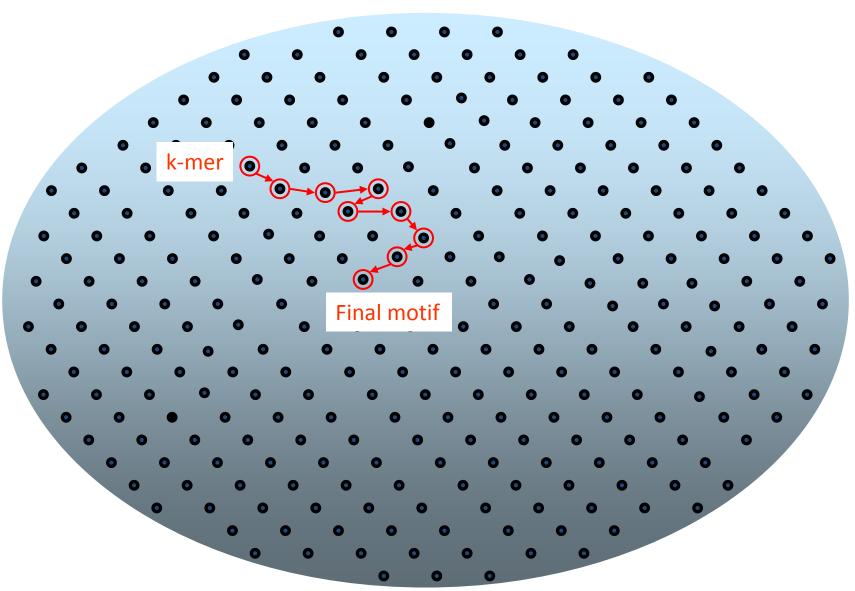
#### A schematic view of the optimization process



#### A schematic view of the optimization process



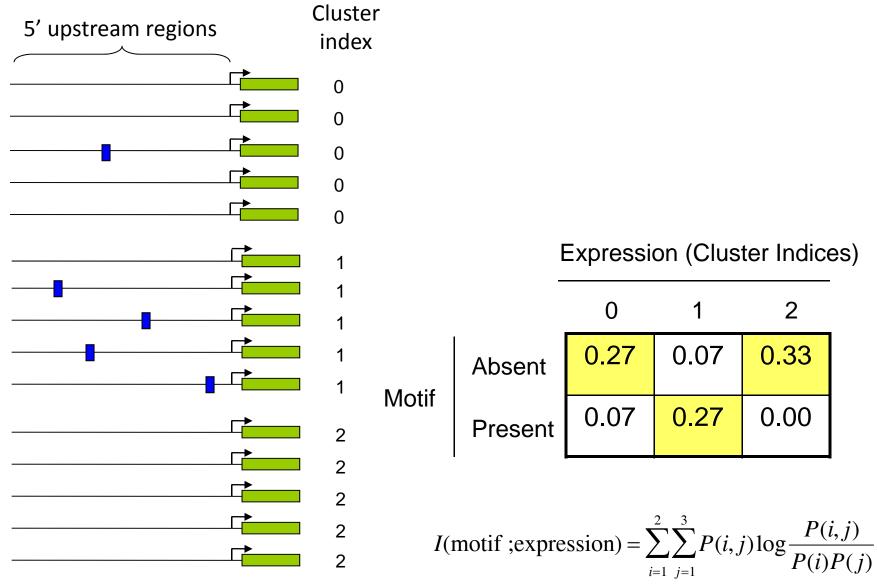
#### A schematic view of the optimization process





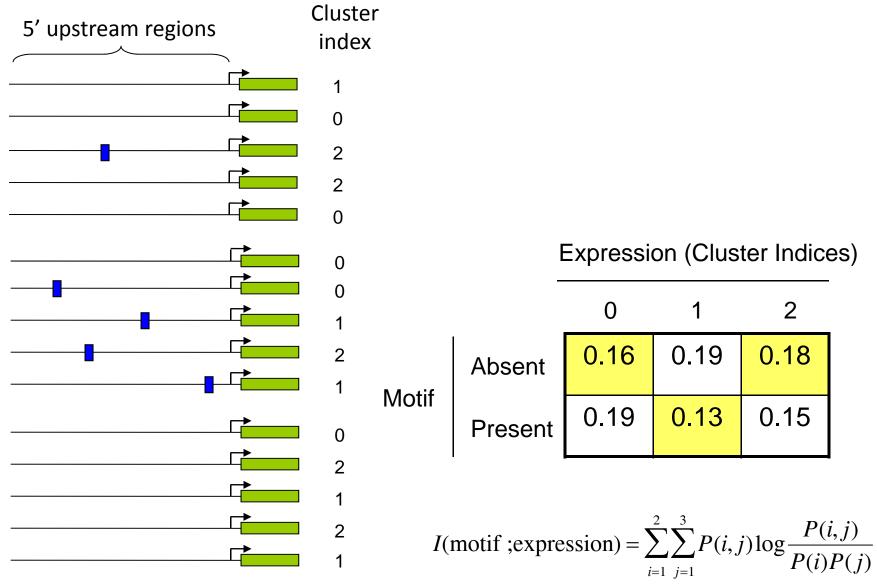
# Is a given motif more informative than expected by chance?





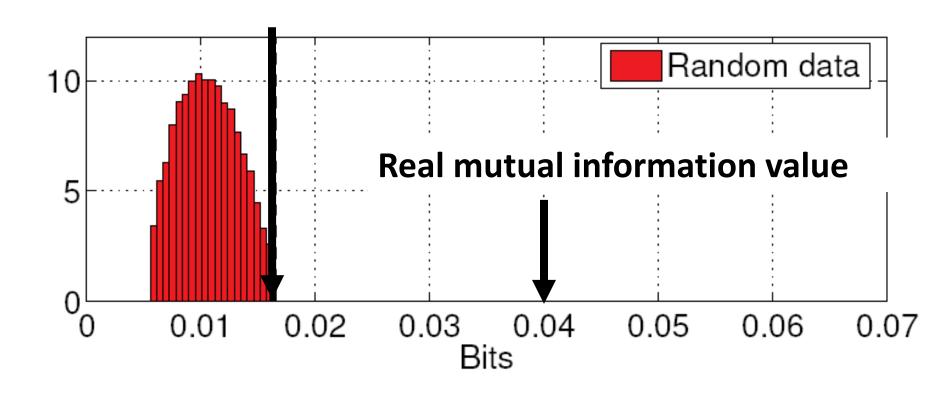
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. . .

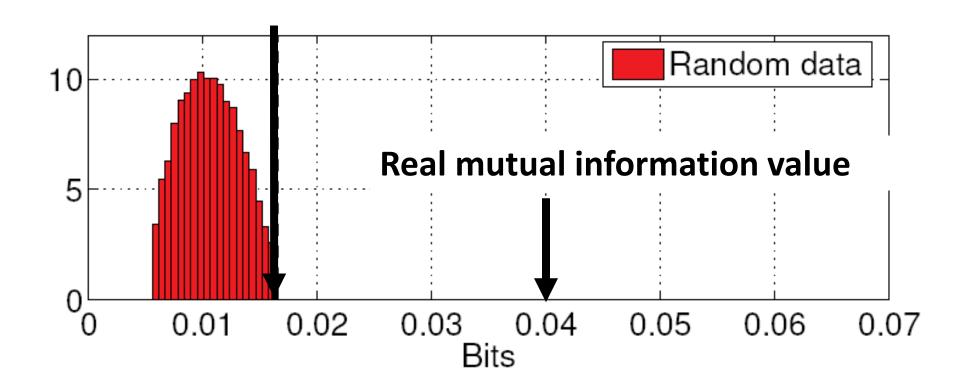
#### Maximum of 10,000 expressionshuffled mutual information values



P-value: probability of obtaining by chance a result at least as extreme as observed result P(X>=x)

Maximum of 10,000 expressionshuffled mutual information values

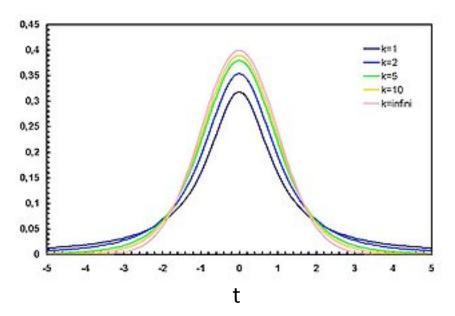
P<10<sup>-4</sup>



### Why non-parametric test?

We don't know what the null distribution of mutual information is like ... depends on sample size, etc.

Null Distribution of T-statistic



Null Distribution of information values

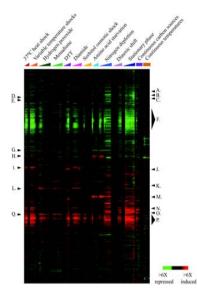


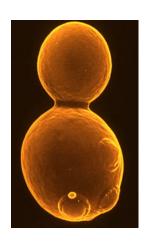
So we estimate it using simulation.



## Yeast stress gene expression program (Gasch et al, 2000)

- 173 microarray conditions
- ~ 5,500 genes
- 80 co-expression clusters
- Runtime ~ 1h (standard PC)





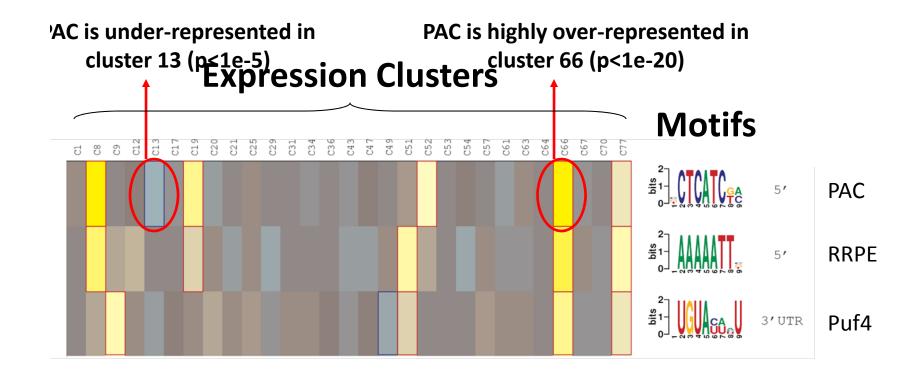
#### **Expression Clusters** # CTCATC<sub>T</sub>e 5. # 5° Z'- SAGU 3'UTR #1-] 3'UTR # COO 5' A S' ACCOLAÇE 5' 3'UTR 3'UTR # 1 FOCACE 5" AC AC # 1 A COLT 6 5' # CA 5' 2 0 00 00 5' I CAACOO 5' 21 c C Ac 5' 21- 1- 10 Vê. 5' 2 COTO 2 3'UTR #1- CAN CV 5' # JUAN 3'UTR #1-C 051

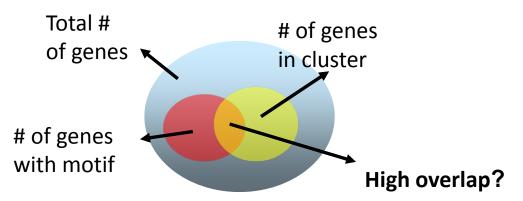
**Predicted** 

**Motifs** 

17 motifs in 5' upstream regions6 motifs in 3'UTRs



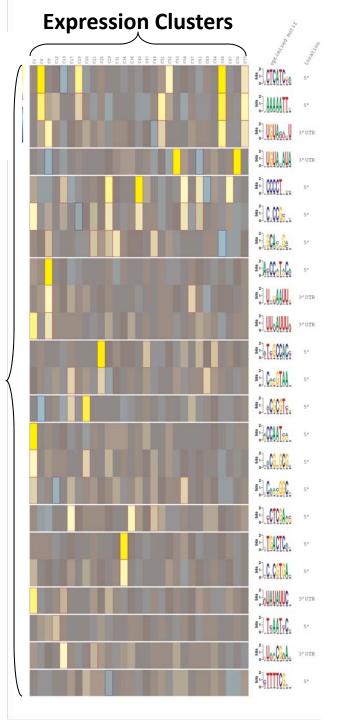




P-value of **over-representation** of a motif in a cluster of genes

$$P(X \ge I) = \sum_{x=I}^{min(s_1,s_2)} \frac{\binom{s_1}{x} \binom{N-s_1}{s_2-x}}{\binom{N}{s_2}}$$

Hypergeometric distribution



17 motifs in 5' upstream regions 6 motifs in 3'UTRs

How many of these motifs are false positives?

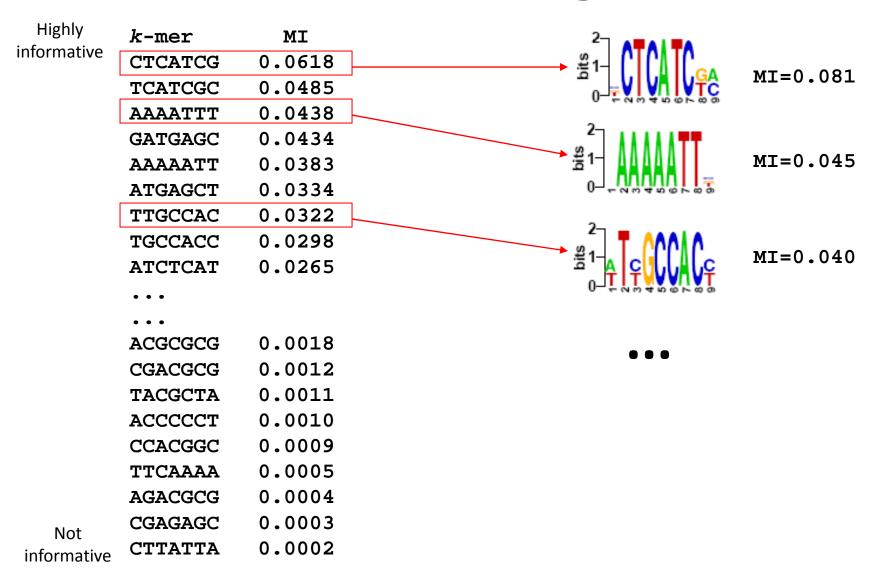
### Predicted Motifs

Where do false positives come from?

- Multiple hypothesis testing (k-mers)
- Overfit by the optimization procedure



## Motif Search Algorithm



# Does the algorithm overfit motifs to the expression?

Cluster 0: 112 genes

Cluster 1: 132 genes — enriched in RAP1 motif

Randomly split cluster 1 into cluster 1A and 1B

Cluster 0: 112 genes

Cluster 1A: 66 genes

Cluster 0 : 112 genes

Cluster 1B: 66 genes

Best motif, MI=0.38 bits

[CT]CC[AG][ACT]AC[AG][CT]

MI=0.301 bits when evaluated

on this dataset

MI=0.29 bits when evaluated on this dataset

Best motif, MI=0.33 bits

[ACG][CT]CC[AG][CT][AG]C[AC]

### Estimating the false discovery rate

- Run motif discovery algorithm (kmers+optimization) on random expression profile
- Count how many motifs we get
- Repeat a large number of times, calculate average number of motifs

#### **Expression Clusters** E- CTCATCAS 5 # 5° Z1- WAG 3'UTR 2 3° UTR 2 000 5° 1 COC 5' 1 C CA 5' E Jemala 5 a'UTR 3'UTR # 1 - COMO 5" Act Act 5 1 A CO C 5' E ACCAN SA # Della 000 E CAACOO 5' 21- COMAC 5' # 5' 2 COTO 2 3'UTR #1- chick 5' #1- JAACOA 3'UTR

**Predicted** 

**Motifs** 

17 motifs in 5' upstream regions 6 motifs in 3'UTRs

~ 0.05 "motifs" when shuffling the gene labels of the clustering partition

### Entropy

$$H(X) = -\sum_{x} P(x) \log P(x)$$

```
X P(X)
0 0.5 H(X)=1 bit
1 0.5
```

```
X P(X)
0 0.8 H(X)=0.72 bits
1 0.2
```

Χ	P(X)	
0	1.0	H(X)=0 bits
1	0.0	

### Mutual Information

$$I(X;Y) = H(Y) - H(Y|X)$$

Uncertainty about Y

the amount of uncertainty remaining about Y after X is known