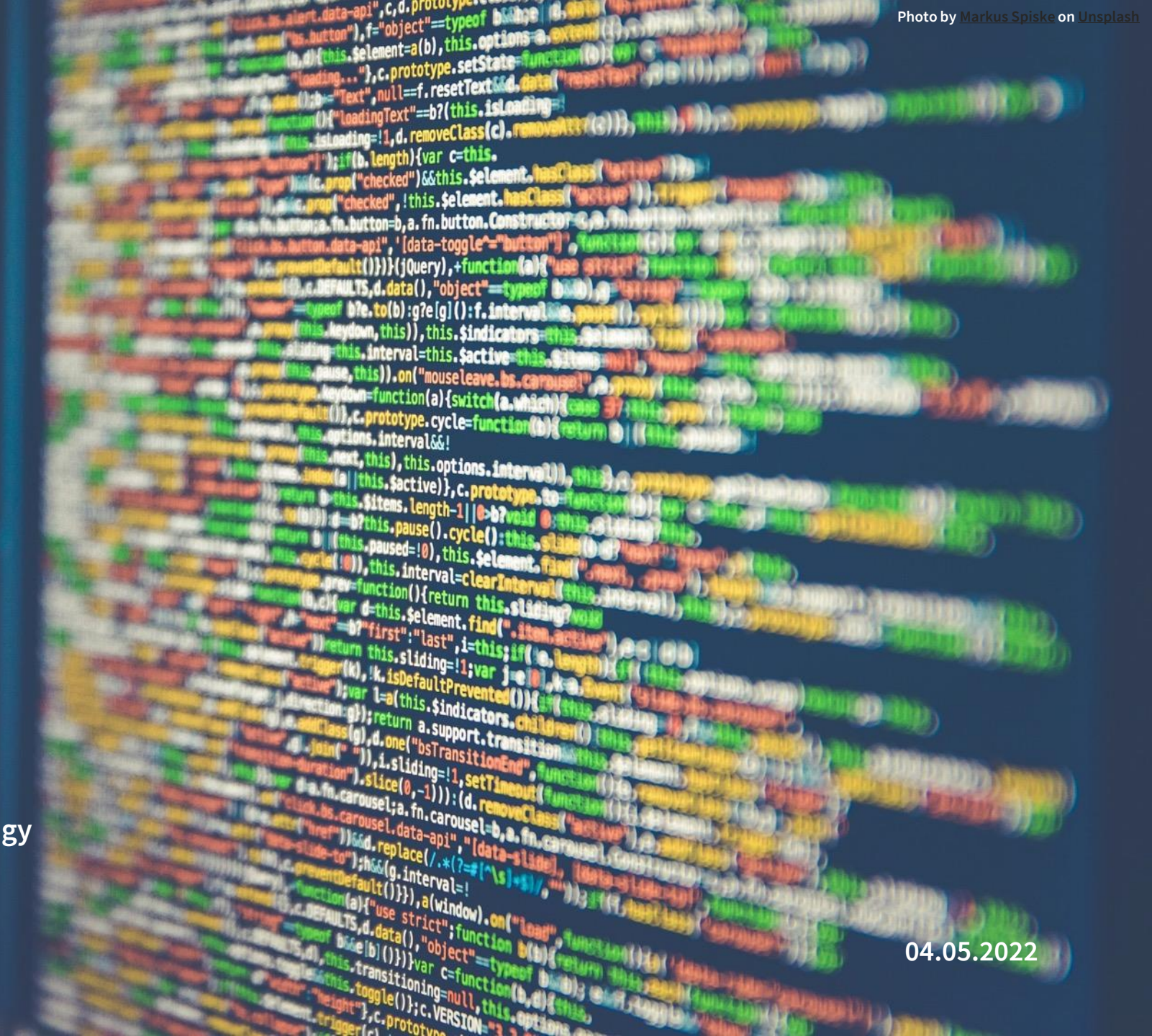


# RNA-Seq for Precision Medicine: translating research into clinical applications


Clinical and Research Genomics  
Spring 2022 Course


Andrea Sboner, PhD (he/him/his)  
Director of Informatics and Computational Biology


 **Weill Cornell Medicine**  
Caryl and Israel Englander  
Institute for Precision Medicine



# What is Precision Medicine?

The right drug... 

...to the right patient ... 

...at the right time! 



# Precision Medicine benefits from advancements in sequencing technology (NGS)

A very brief (and incomplete) history of genomics sequencing

- 3.1 billion nucleotides (DNA) sequenced and assembled (chr1, chr2, ..., chr22, chrX, chrY) from a few individuals
- Costs: \$3.8B → \$796B economic impact
- 1990 – 2003 (13 years)

## Economic Impact of the Human Genome Project

*How a \$3.8 billion investment drove \$796 billion in economic impact, created 310,000 jobs and launched the genomic revolution*

Simon Tripp and Martin Grueber  
Battelle Memorial Institute  
Technology Partnership Practice  
May 2011

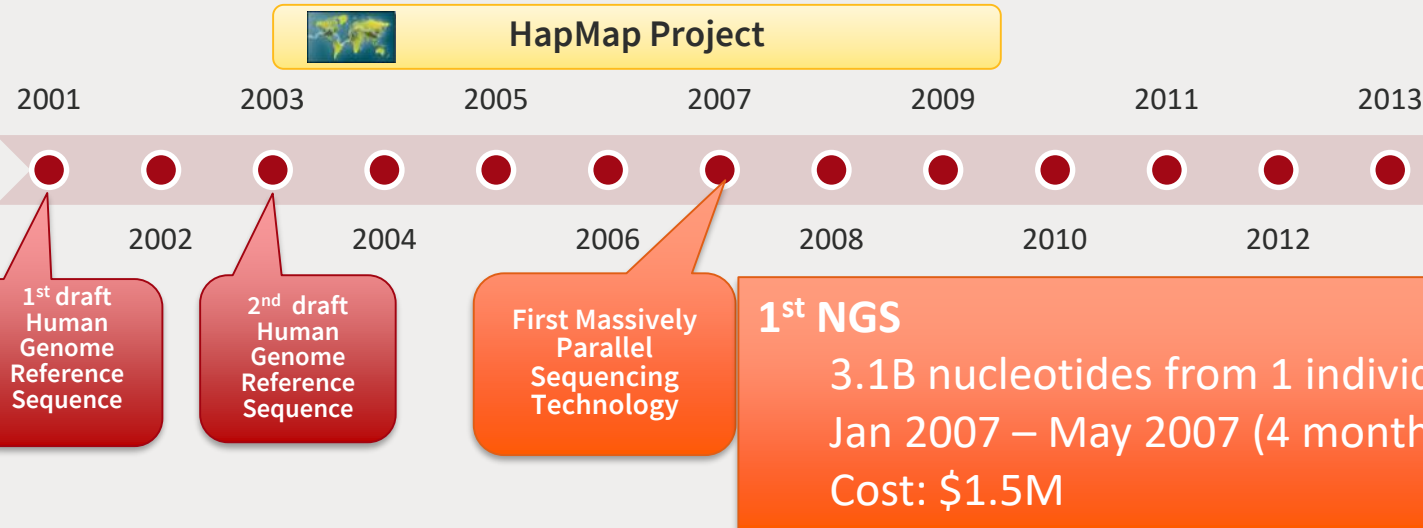
nature

Vol 452 | 17 April 2008 | doi:10.1038/nature06884

## LETTERS

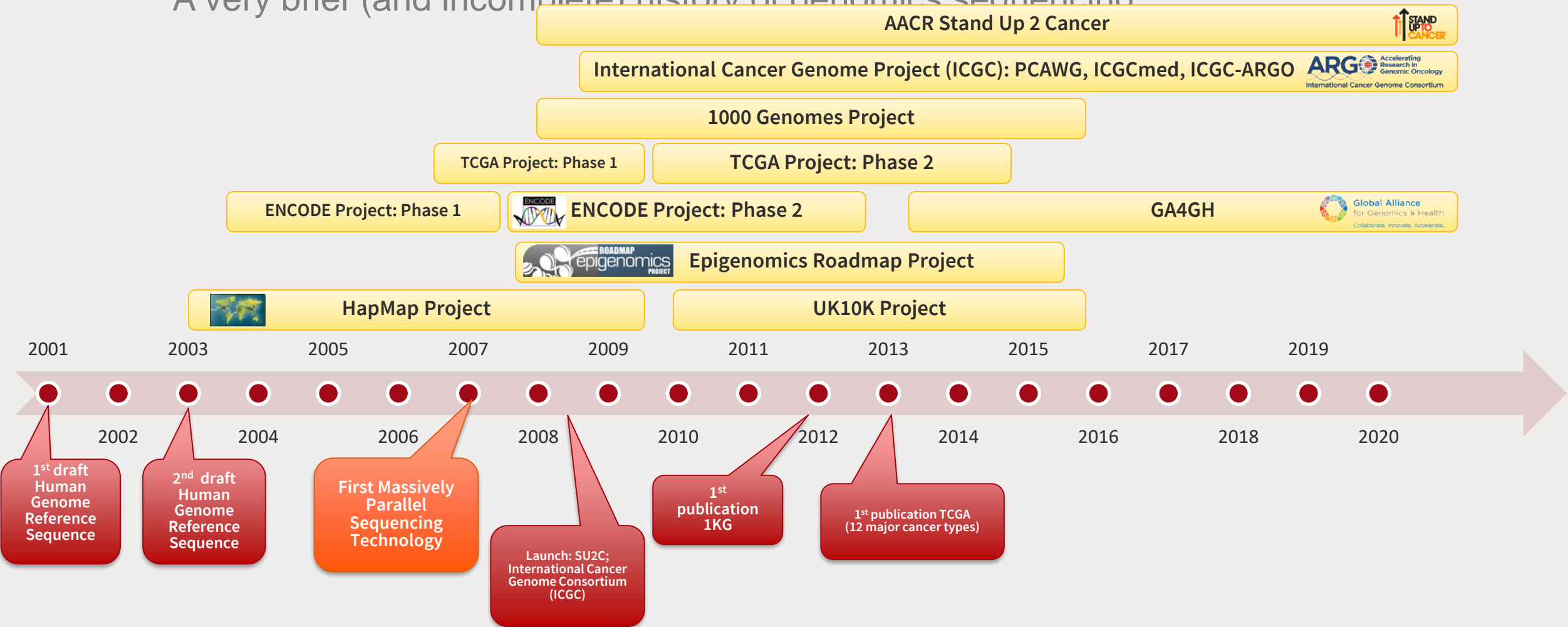
### The complete genome of an individual by massively parallel DNA sequencing

David A. Wheeler<sup>1\*</sup>, Maithreyan Srinivasan<sup>2\*</sup>, Michael Egholm<sup>2\*</sup>, Yufeng Shen<sup>1\*</sup>, Lei Chen<sup>1</sup>, Amy McGuire<sup>3</sup>, Wen He<sup>2</sup>, Yi-Ju Chen<sup>2</sup>, Vinod Makhijani<sup>2</sup>, G. Thomas Roth<sup>2</sup>, Xavier Gomes<sup>2</sup>, Karrie Tartaro<sup>2†</sup>, Faheem Niazi<sup>2</sup>, Cynthia L. Turcotte<sup>2</sup>, Gerard P. Irzyk<sup>2</sup>, James R. Lupski<sup>4,5,6</sup>, Craig Chinault<sup>4</sup>, Xing-zhi Song<sup>1</sup>, Yue Liu<sup>1</sup>, Ye Yuan<sup>1</sup>, Lynne Nazareth<sup>1</sup>, Xiang Qin<sup>1</sup>, Donna M. Muzny<sup>1</sup>, Marcel Margulies<sup>2</sup>, George M. Weinstock<sup>1,4</sup>, Richard A. Gibbs<sup>1,4</sup> & Jonathan M. Rothberg<sup>2†</sup>

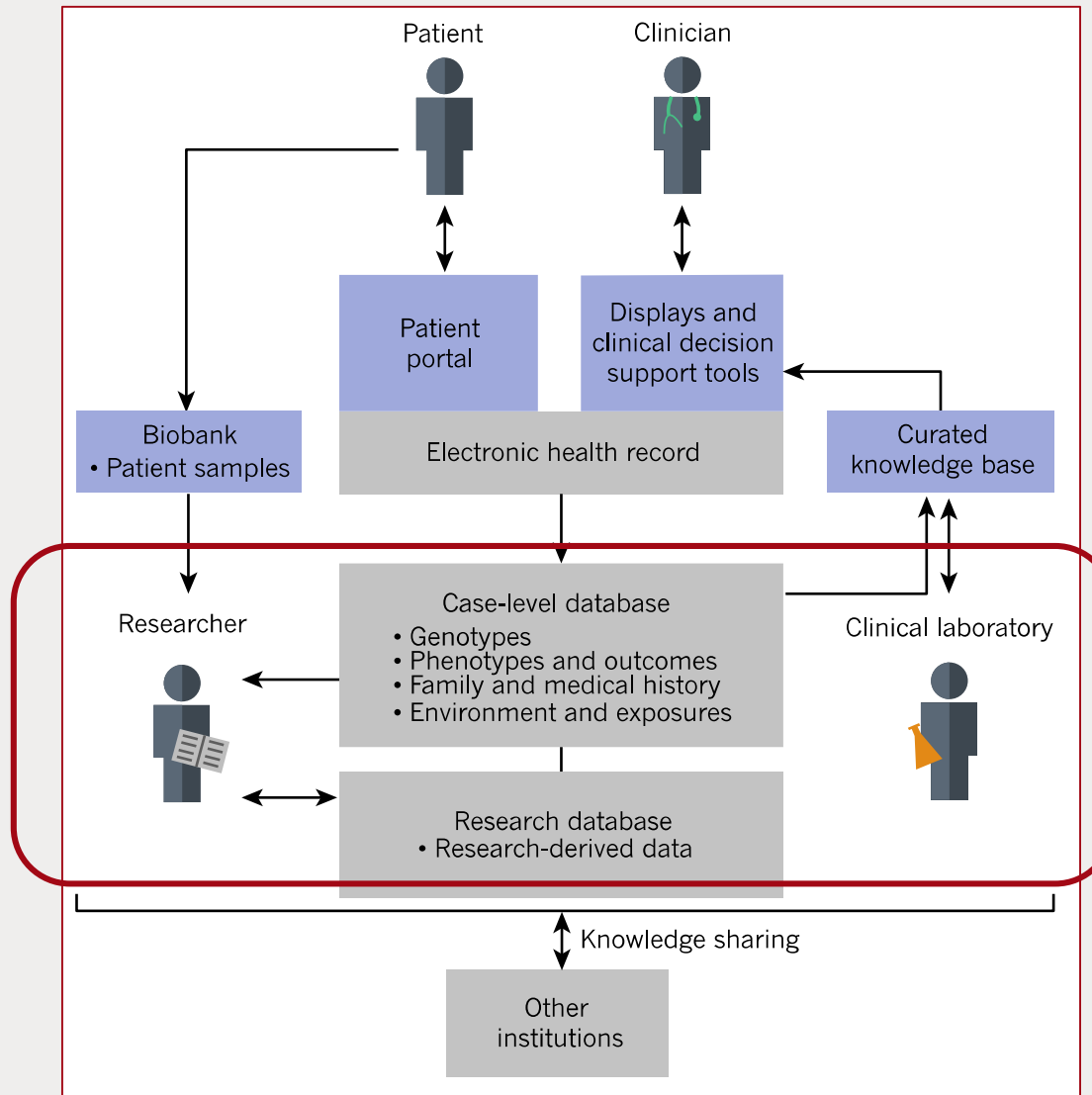


# Precision Medicine benefits from advancements in sequencing technology (NGS)

A very brief (and incomplete) history of genomics sequencing

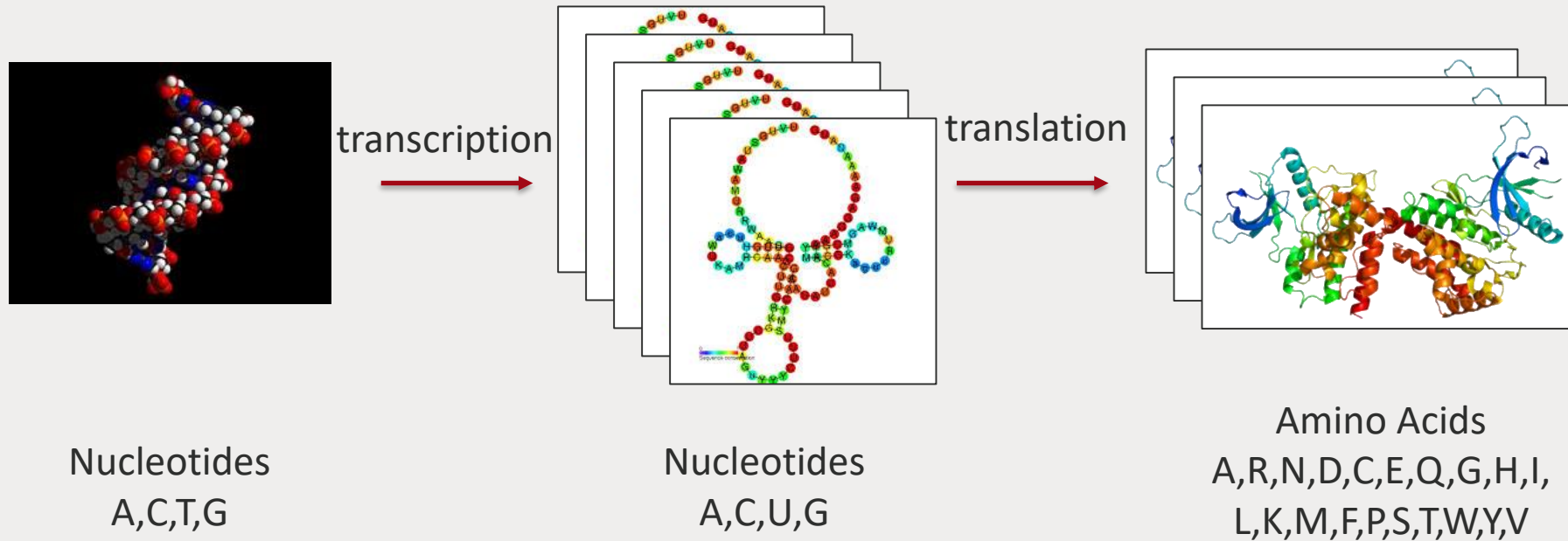


# The Precision Medicine Ecosystem



S. J. Aronson, H. L. Rehm, *Nature*. **526**, 336–342 (2015).

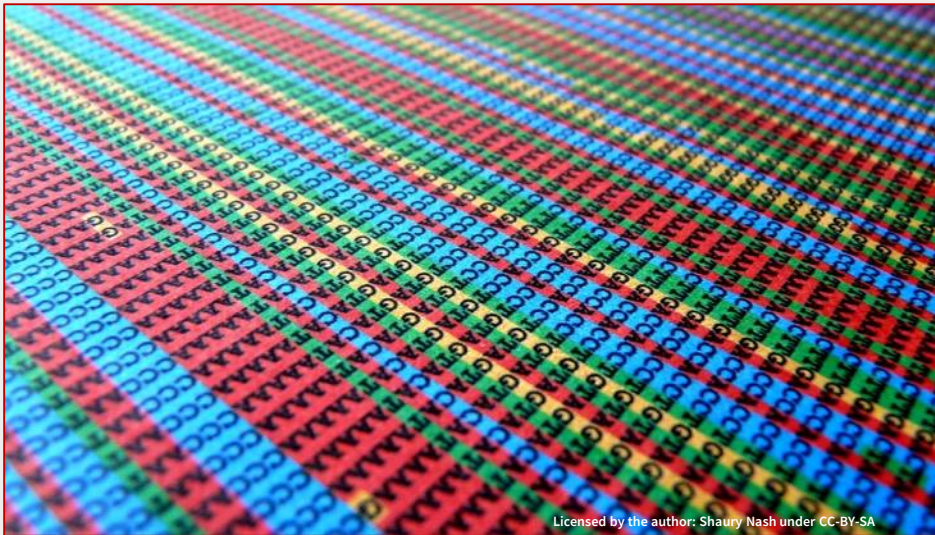
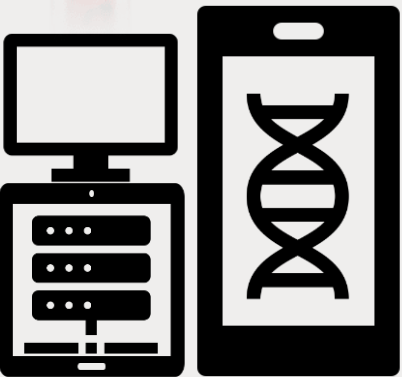
# Transcriptome profiling



Transcriptome profiling goal is to characterize the RNA in a tissue or cell.

The 'simpler' structure of RNA allows to employ most techniques used for DNA analysis – hybridization, polymerase chain reaction, etc.

# Sequencing & Analysis: A translation of biological material into a common language



## Analysis

Deletion



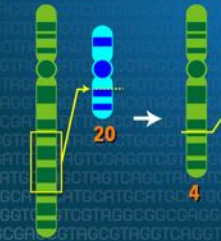
Duplication



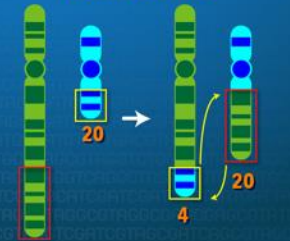
Inversion



Substitution



Translocation



## SNVs & InDels

Tumor: ATTCG**A**TTATTC  
Reference: ATTCGCTTATTC

Tumor: ATTCG**AACT**TTATTC  
Reference: ATTCG**---**TTATTC

Tumor: ATTCG**---**TATTC  
Reference: ATTCGCTTATTC

# Genome Era (1990s – 2000s)

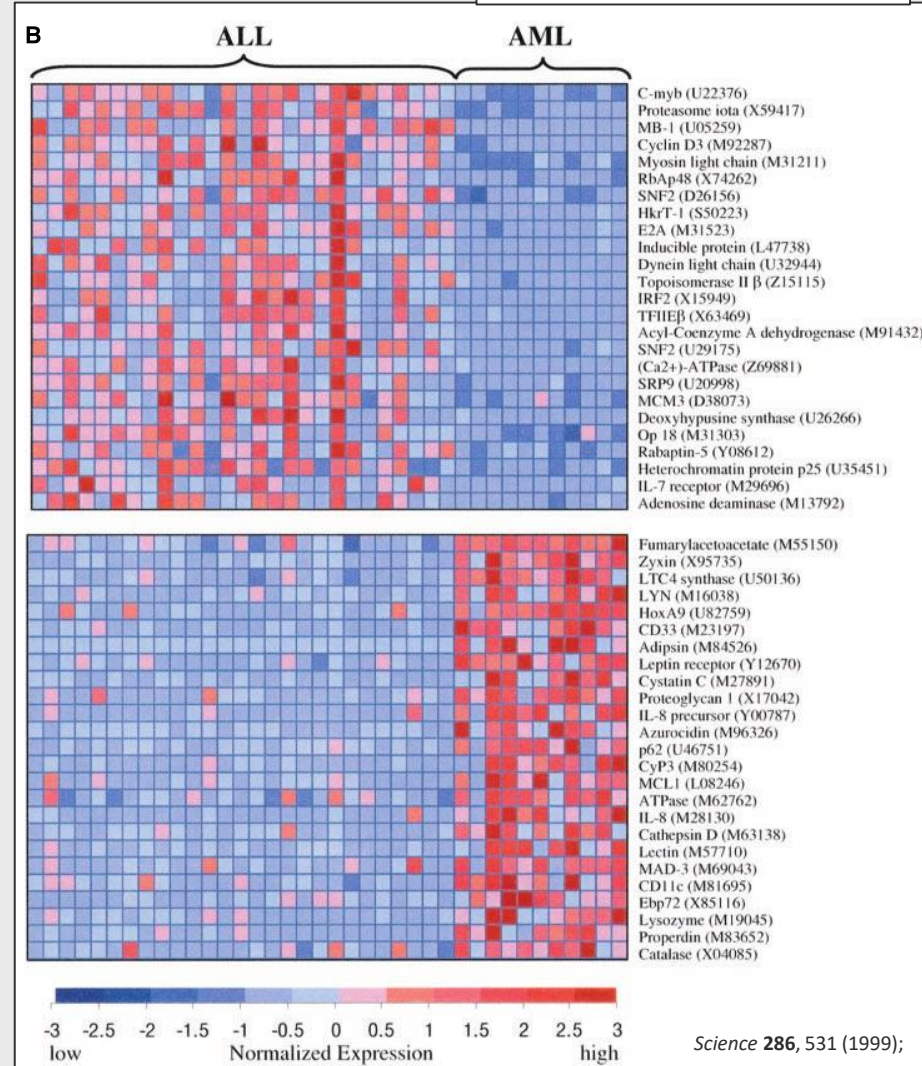
~ 1991 Expressed Sequence Tags (ESTs) sequencing (500-800 nucleotides)

~ 1995 Series Analysis of Gene Expression (SAGE) (9-12 nucleotides)

~ 1999 Microarray

## Molecular Classification of Cancer: Class Discovery and Class Prediction by Gene Expression Monitoring

T. R. Golub,<sup>1,2\*</sup> D. K. Slonim,<sup>1†</sup> P. Tamayo,<sup>1</sup> C. Huard,<sup>1</sup> M. Gaasenbeek,<sup>1</sup> J. P. Mesirov,<sup>1</sup> H. Coller,<sup>1</sup> M. L. Loh,<sup>2</sup> J. R. Downing,<sup>3</sup> M. A. Caligiuri,<sup>4</sup> C. D. Bloomfield,<sup>4</sup> E. S. Lander<sup>1,5\*</sup>





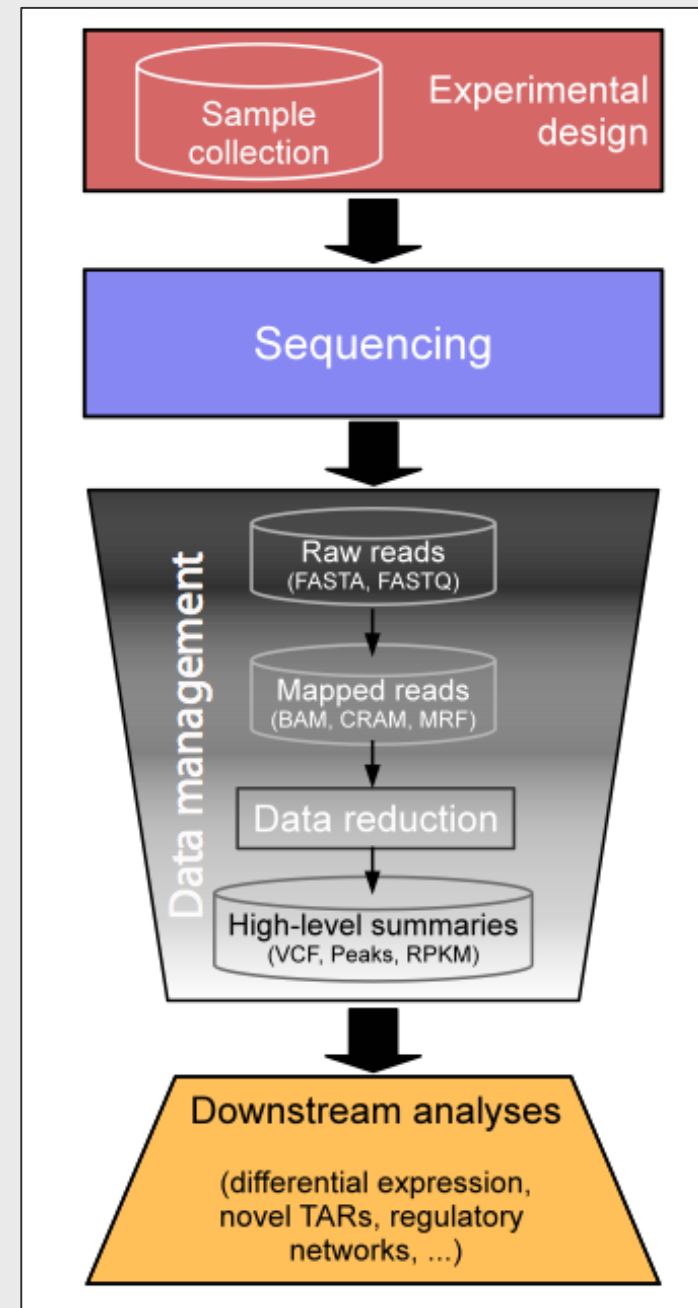
# RNA-Seq Experiment

Data management:

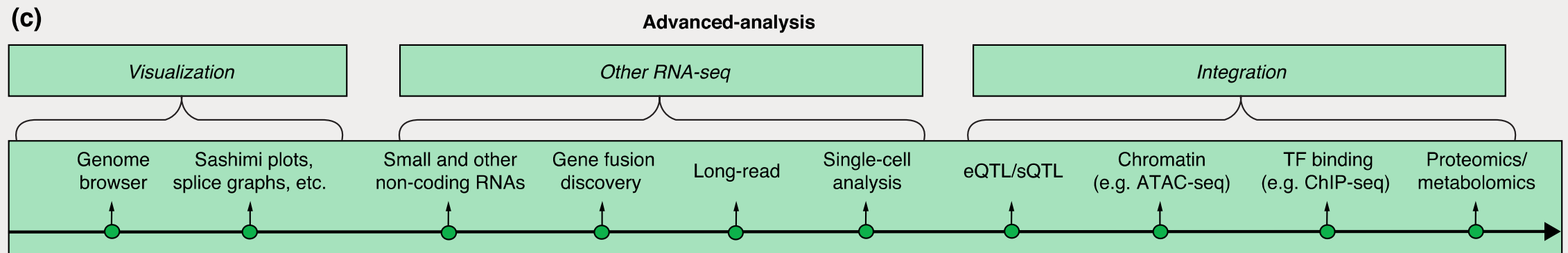
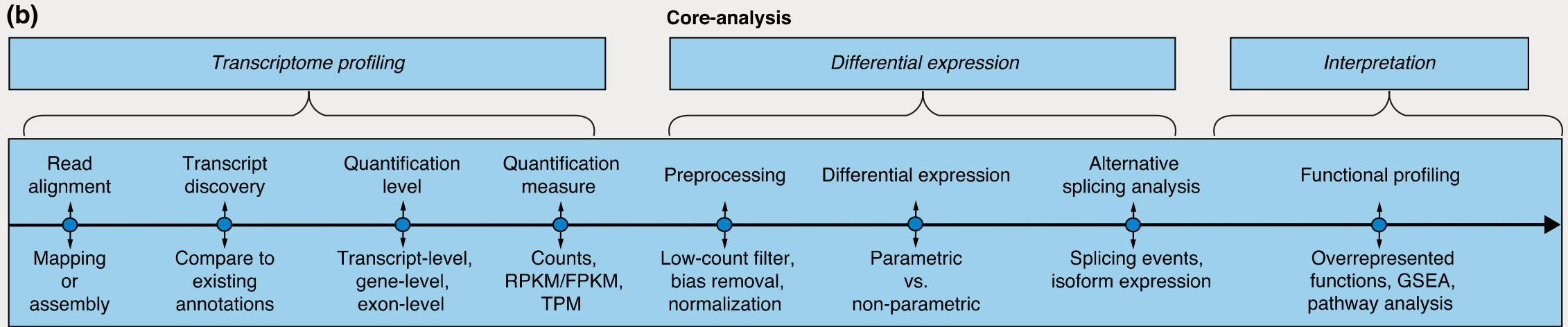
Mapping the reads

Creating summaries

Downstream analysis: *the interesting stuff*  
Differential expression, chimeric transcripts,  
novel transcribed regions, etc.

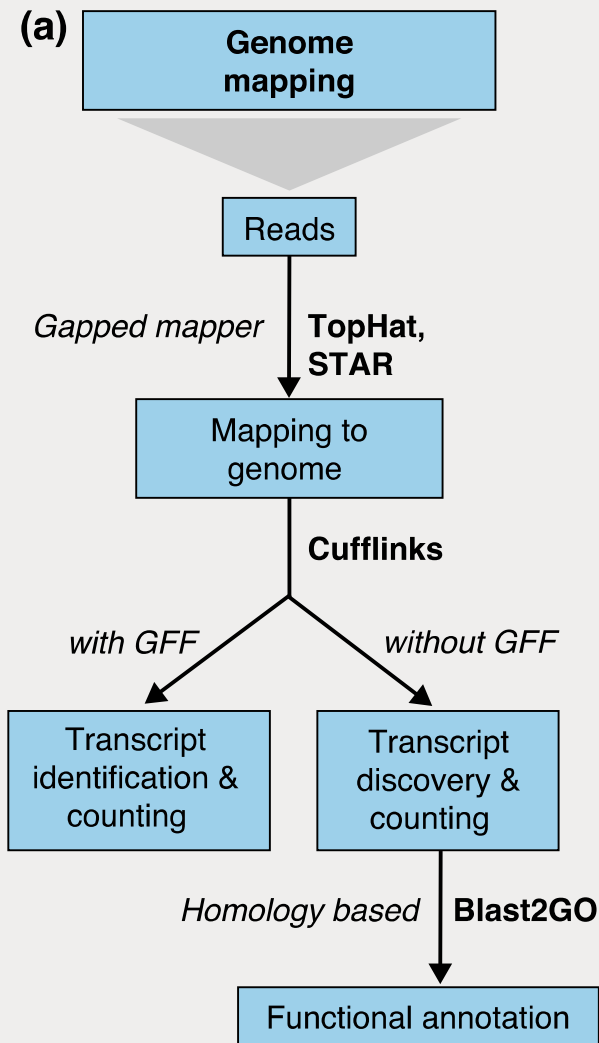


# Roadmap for RNA-seq analyses



A. Conesa et al., *Genome Biology*. 17, 13 (2016).

# Alignments for Transcriptomics (RNA-seq)



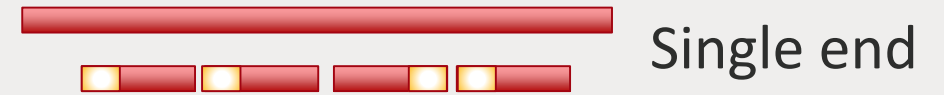
A. Conesa et al., *Genome Biology*. **17**, 13 (2016).

# Expression Quantification

FPKM/RPKM: Fragment/Reads per Kilobase of exonic region per Million of reads

TPM: transcripts per million

Normalization strategies affect results of comparisons (ERCC-spike-ins)

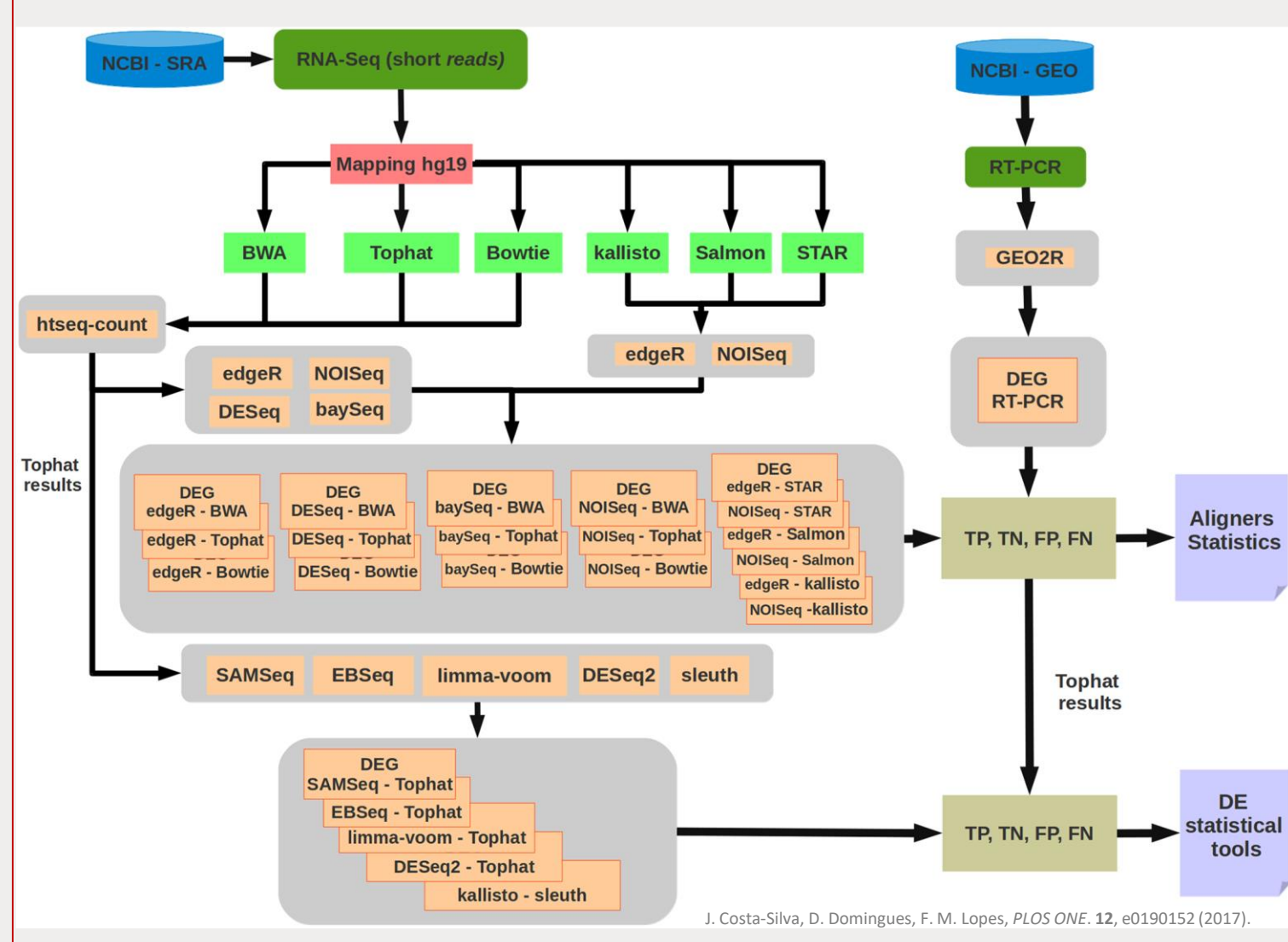
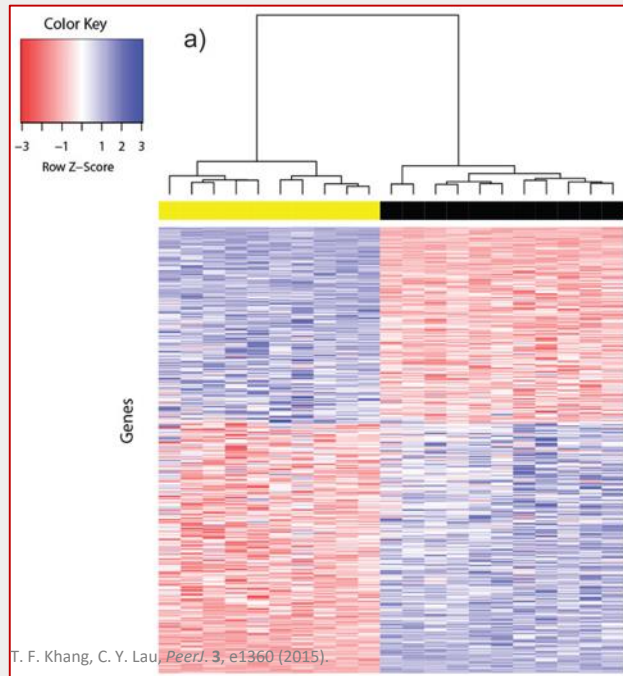


1. Divide the total # reads by 1M=scaling factor
2. Divide the read/fragment counts by the scaling factor=RPM/FPM
3. Divide RPM by the length of the genes in KB=RPKM/FPKM

1. Divide the read/fragment counts by the length of the gene=RPK
2. Sum all RPK and divide by 1M=scaling factor
3. Divide RPK by the scaling factor=TPM

# Differential Expression

Comparison of groups



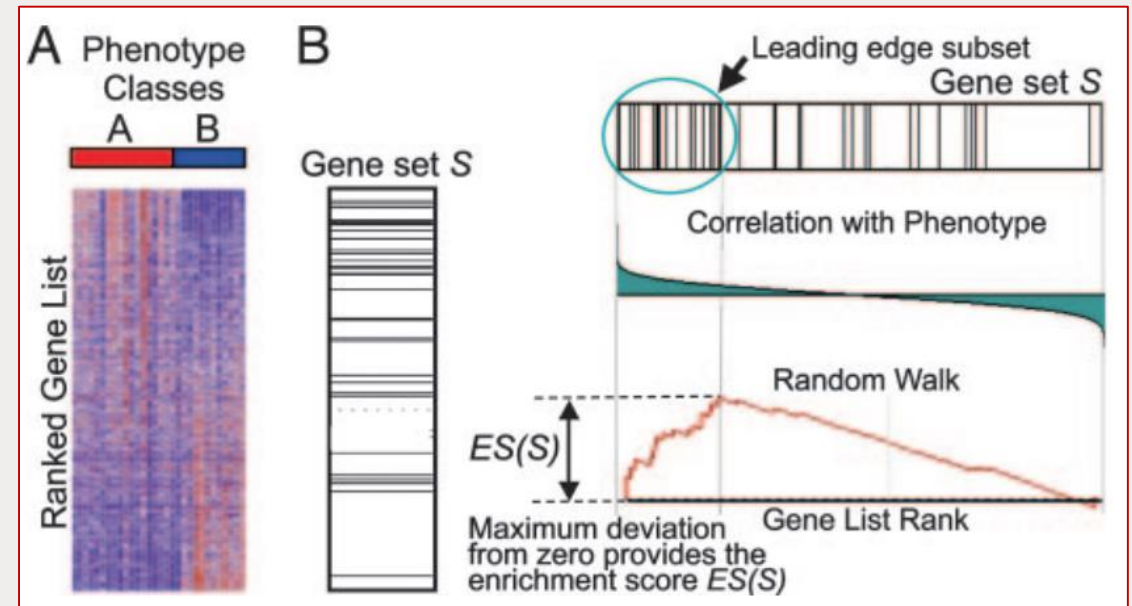
We have identified that the impact of the mapping tool on the final results is minimal, indicating the DEGs identification method is the main choice for differential expression analysis in RNA-Seq data.

We *did not* identify among the evaluated methods a tool that obtained *optimum results in all performance measures*, for the evaluated experimental conditions. The NOIseq, DESeq2 and limma+vomm methods present the best individual results with 95%, 95% and 93% of Specificity and 80%, 84% and 81% of True Positive Rate, respectively.

# Pathway analysis (gene set enrichment)

Are a group of genes dys-regulated in a certain condition?

“The basic assumption is that although large changes in individual genes can have significant effects on pathways, weaker but coordinated changes in sets of functionally related genes (i.e., pathways) can also have significant effects. Therefore, the gene-level statistics for all genes in a pathway are aggregated into a single pathway-level statistic (e.g. the sum of all log-fold changes), which will then be evaluated.”



Introduction to differential gene expression analysis using RNA-seq Written by Friederike Dundar, Luce Skrabanek, Paul Zumbo <https://chagall.med.cornell.edu/RNASEQcourse/Intro2RNAseq.pdf>

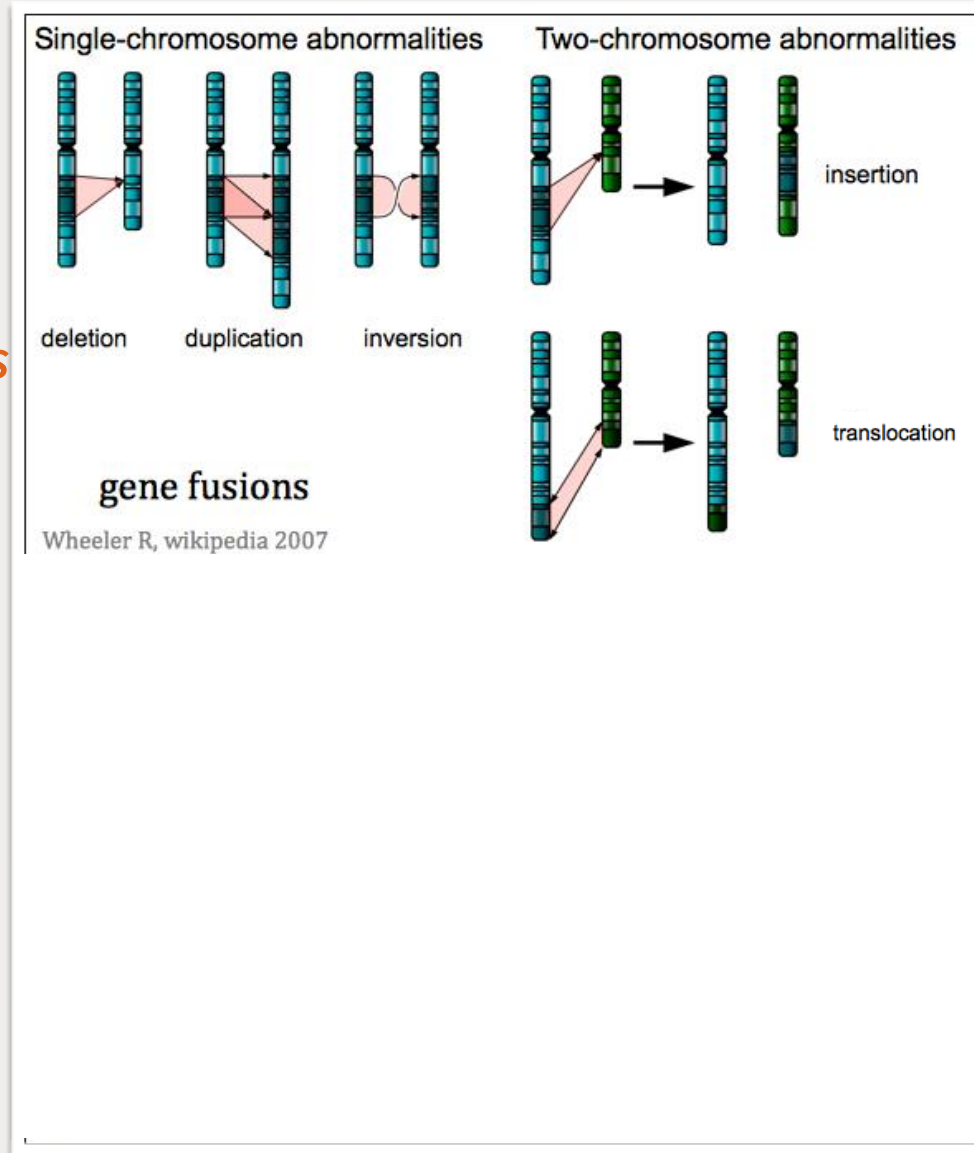
# What are chimeric transcripts?

Transcripts that are *not co-linear* in the genome space

They can arise from:

genomic rearrangements, i.e. *gene fusions*

post-transcriptional events, i.e. *trans-splicing or cis-splicing*



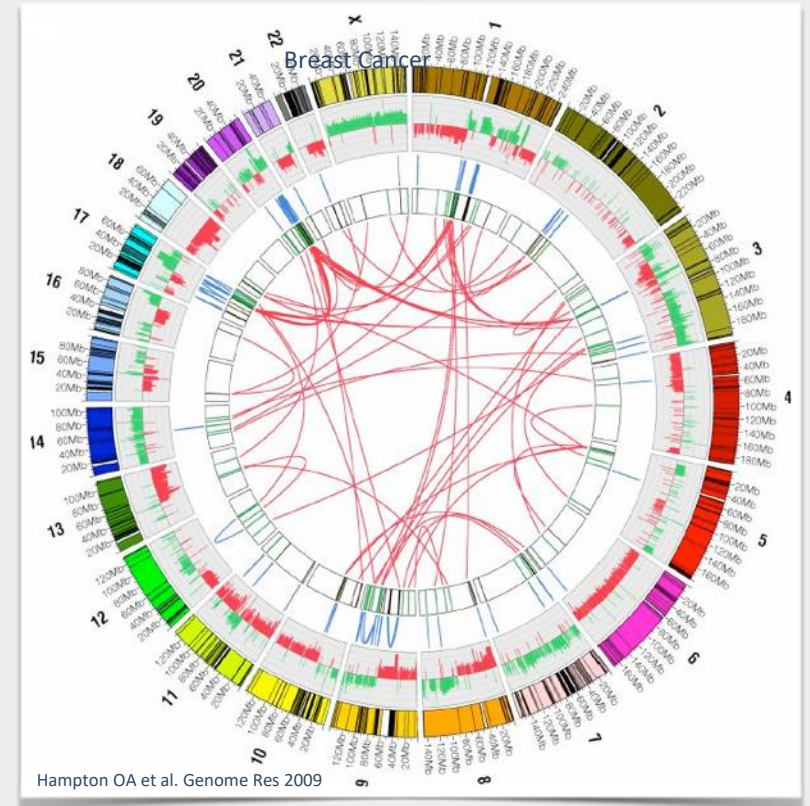
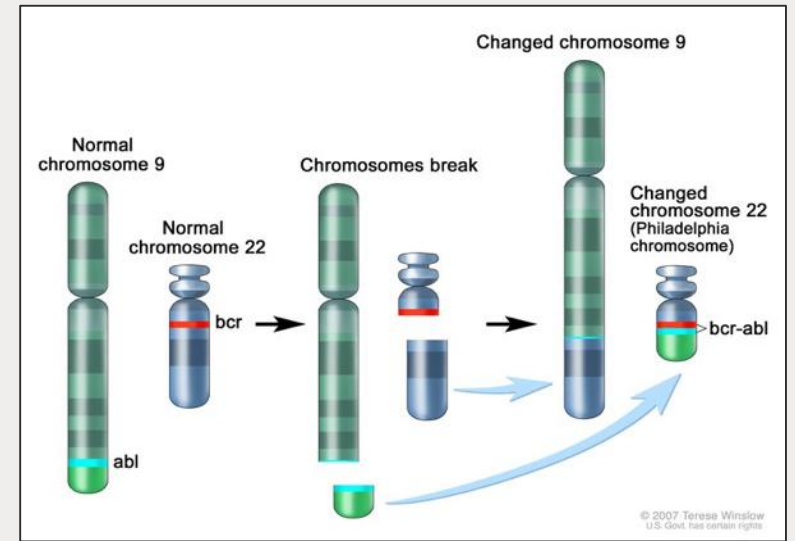
# Why are they (gene fusions) important?

Fusion genes are often *oncogenes*

Ex: BCR-ABL1 (Philadelphia chromosome) in Chronic myelogenous leukemia (CML) and Acute Lymphoblastic leukemia (ALL)  $t(9;22)(q34;q11)$

Fusion involving a proto-oncogene with a strong promoter resulting in *upregulation* (lymphomas)

Ex: (IgH locus)-MYC in Burkitt's lymphoma (cMYC over-expressed)





# Why are they (trans-splicing events) important?

Trans(cis)-splicing was initially found in lower eukariotes, such as trypanosomes and worms

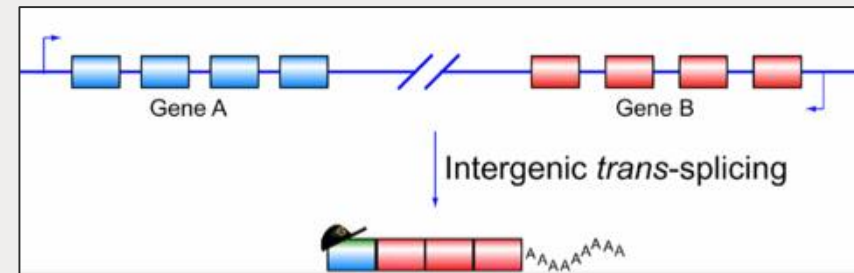
Short sequences of nucleotides are trans-spliced to distant 5' of many protein coding genes

Recently, they were found in mammalian cells:

JAZF1-SUZ12 in endometrial stroma cells (Li et al. Science 2008)

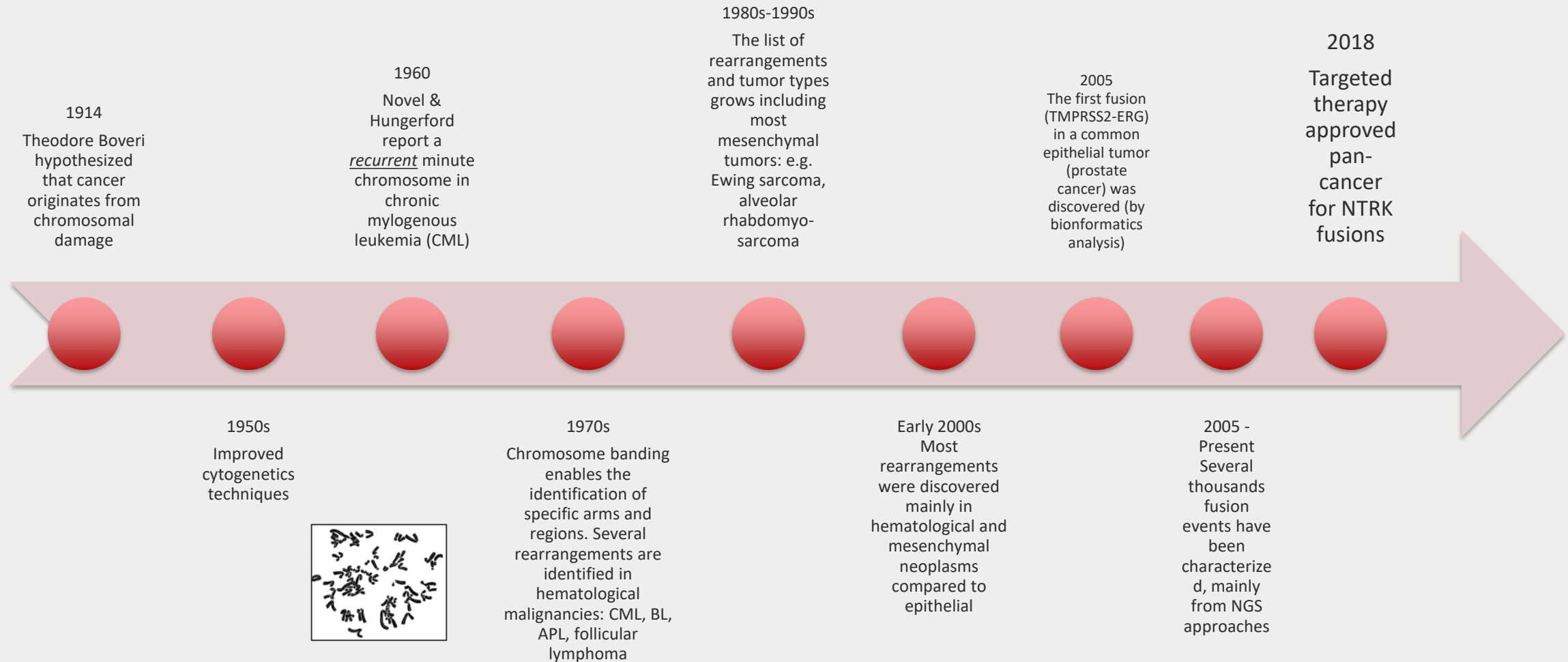
SLC45A3-ELK4 in prostate tissues (Rickman et al. Cancer Res 2009)

65% of protein-coding genes have distal 5' transcription start sites (ENCODE pilot) --> revised to ~50% the ENCODE 2012



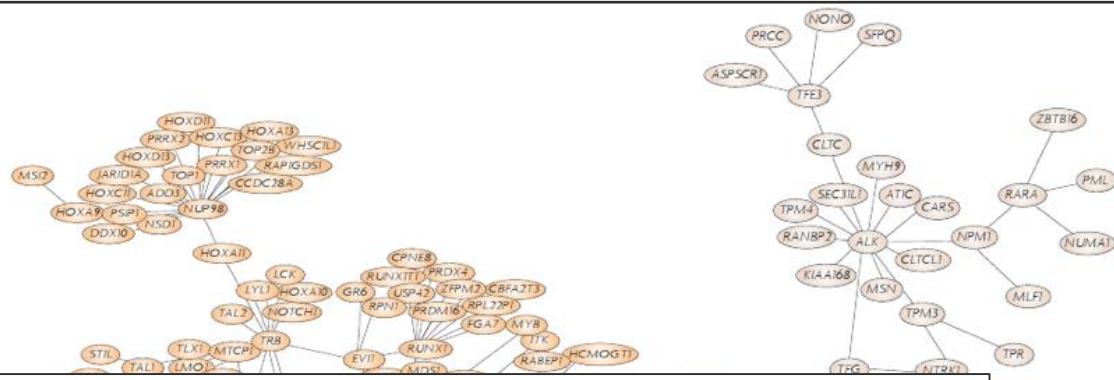
Horiuchi, Takayuki, and Toshiro Aigaki. Biology of the Cell 98, no. 2 (January 9, 2012): 135–140.

# An historical perspective of gene fusions

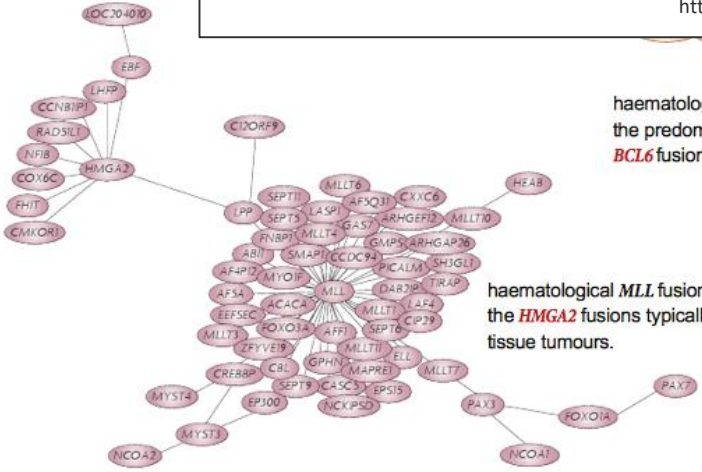


# How many different gene fusions do we know?

- 358 gene fusion
- 337 different genes
- ~90% form three clusters



National Cancer Institute  
**CANCER GENOME ANATOMY PROJECT**  
 Cases: 72,105  
 Fusions: 32,795  
 Mitelman Database of Chromosome Aberrations and Gene Fusions in Cancer (2022). Mitelman F, Johansson B and Mertens F (Eds.), <https://mitelmandatabase.isb-cgc.org>



haematological *ETV6*, *IGH* and *NUP98* fusions the predominantly lymphoma-associated *BCL6* fusions as well as epithelial *RET* fusions.

haematological *MLL* fusions connected to the *HMGA2* fusions typically found in soft tissue tumours.

lymphoma-associated *ALK* fusions, the carcinoma-associated transcription factor for IGHM enhancer 3 (*TFE3*) fusions, and the sarcoma-associated *EWSR1* fusions.

Mitelman F et al, Nature Rev Cancer 2007

# Gene fusions are important for clinical treatment



Open Access Review

**ESMO Open** *Cancer Horizons* **NTRK gene fusions as novel targets of cancer therapy across multiple tumour types**

CrossMark

Alessio Amatu,<sup>1</sup> Andrea Sartore-Bianchi,<sup>1</sup> Salvatore Siena<sup>1,2</sup>

Amatu A, et al. ESMO Open 2016;1:e000023. doi:10.1136/esmoopen-2015-000023

FDA U.S. FOOD & DRUG ADMINISTRATION

Home Food Drugs Medical Devices Radiation-Emitting Products Vaccines, Blood & Biologics Animal & Veterinary Cosmetics Tobacco Products

### Drugs

Home > Drugs > Drug Approvals and Databases > Approved Drugs

#### Approved Drugs

Hematology/Oncology (Cancer) Approvals & Safety Notifications

Drug Information Soundcast in Clinical Oncology (D.I.S.C.O.)

Approved Drug Products with Therapeutic Equivalence Evaluations (Orange Book)

## FDA approves larotrectinib for solid tumors with NTRK gene fusions

SHARE TWEET LINKEDIN PINTEREST EMAIL PRINT

Listen to the FDA D.I.S.C.O. podcast about this approval

On November 26, 2018, the Food and Drug Administration granted accelerated approval to larotrectinib (VITRAKVI, Loxo Oncology Inc. and Bayer) for adult and pediatric patients with solid tumors that have a neurotrophic receptor tyrosine kinase (NTRK) gene fusion without a known acquired resistance mutation, that are either metastatic or recurrent and unresectable.

## FDA approves two cancer treatments after expedited reviews

By Naomi Thomas, CNN  
Updated 6:25 PM ET, Wed November 28, 2018

HERE'S TO YOUR HEALTH

FDA APPROVES GROUNDBREAKING NEW CANCER DRUG

More from CNN

- Two Pitts teenager a toddler
- Photogra selfie eve years
- Meghan Markle Makes First Public

# ... and diagnostic/prognostic purposes


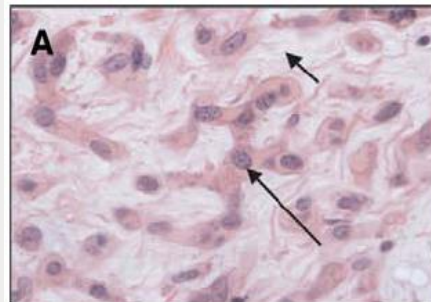
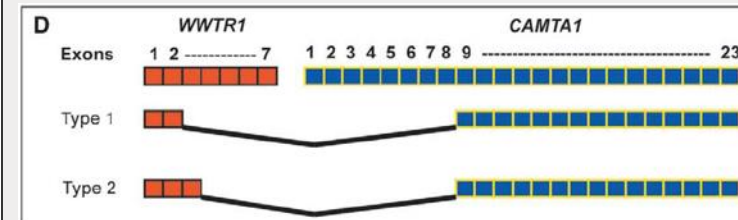
RESEARCH ARTICLE

CANCER

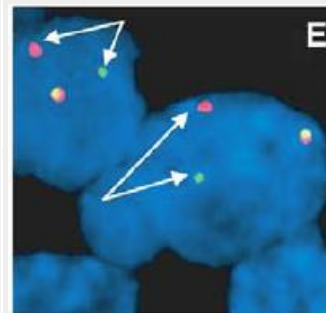
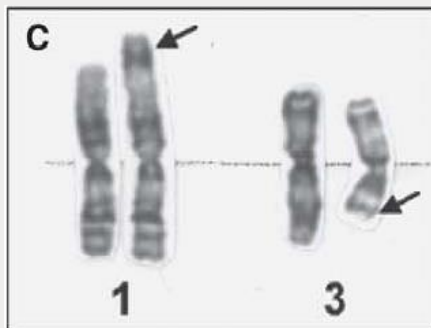
## Identification of a Disease-Defining Gene Fusion in Epithelioid Hemangioendothelioma

Munir R. Tanas,<sup>1</sup> Andrea Sboner,<sup>2</sup> Andre M. Oliveira,<sup>3</sup> Michele R. Erickson-Johnson,<sup>3</sup> Jessica Hespelt,<sup>1</sup> Philip J. Hanwright,<sup>1</sup> John Flanagan,<sup>4</sup> Yuling Luo,<sup>4</sup> Kerry Fenwick,<sup>5</sup> Rachael Natrajan,<sup>5</sup> Costas Mitsopoulos,<sup>5</sup> Marketa Zvelebil,<sup>5</sup> Benjamin L. Hoch,<sup>6</sup> Sharon W. Weiss,<sup>7</sup> Maria Debiec-Rychter,<sup>8</sup> Raf Scot,<sup>9</sup> Rob B. West,<sup>10</sup> Alexander J. Lazar,<sup>11</sup> Alan Ashworth,<sup>5</sup> Jorge S. Reis-Filho,<sup>5</sup> Christopher J. Lord,<sup>5</sup> Mark B. Gerstein,<sup>2,12</sup> Mark A. Rubin,<sup>13</sup> Brian P. Rubin<sup>1\*</sup>

www.ScienceTranslationalMedicine.org 31 August 2011 Vol 3 Issue 98 98ra82

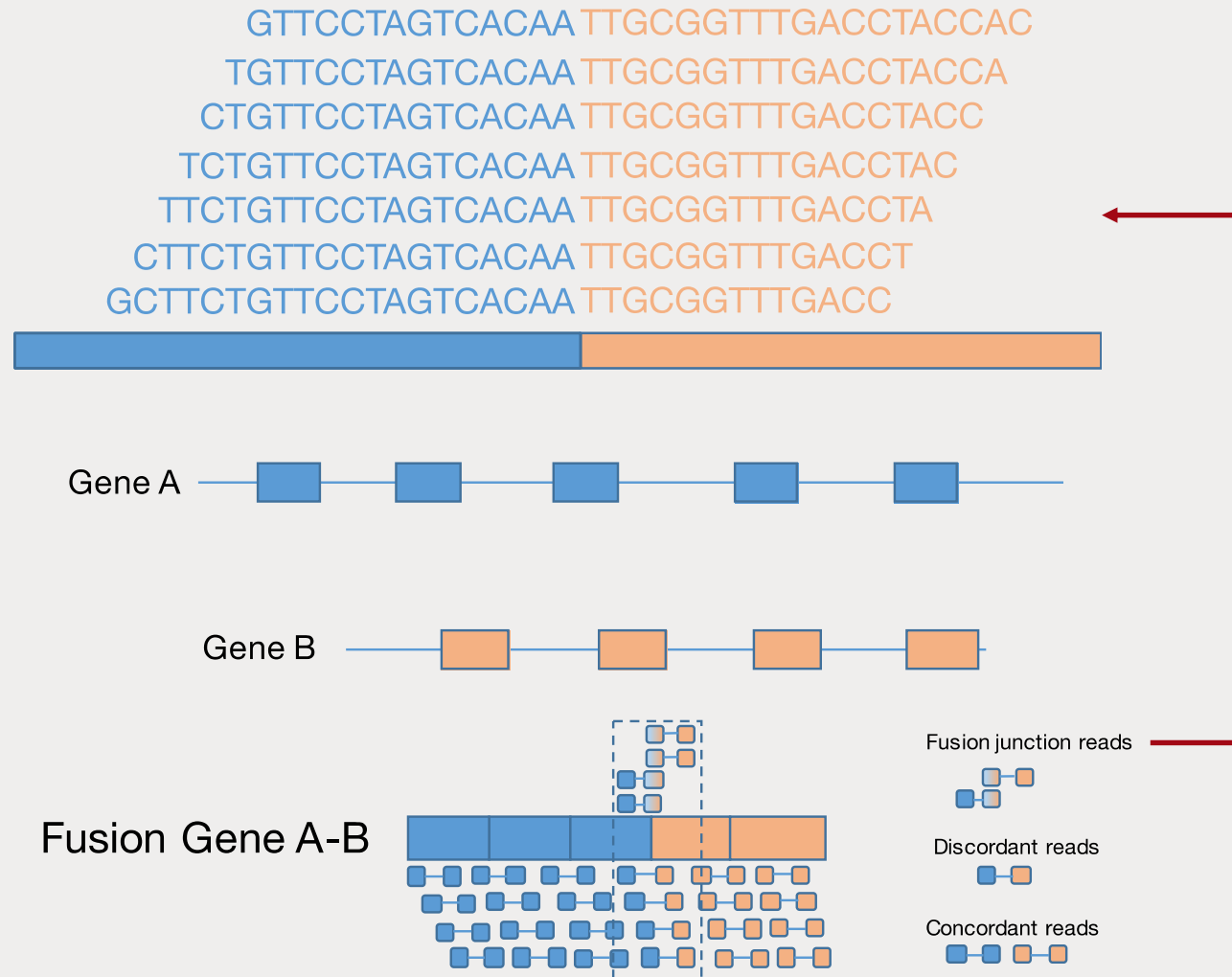



Exclusively present in  
*epithelioid*  
*hemangioendothelioma*



G	WWTR1		CAMTA1	
	Positive /total	%	Positive /total	%
Epithelioid hemangioendothelioma	42/47	89%	39/45	87%
Angiosarcoma, NOS	0/42	0%	0/35	0%
Epithelioid angiosarcoma	0/7	0%	0/7	0%
Intimal sarcoma	0/5	0%	0/3	0%
Kaposi's sarcoma	0/4	0%	0/4	0%
Malignant hemangioendothelioma, NOS	0/1	0%	0/1	0%
Retiform hemangioendothelioma	0/1	0%	0/1	0%
Kaposiform hemangioendothelioma	0/3	0%	0/2	0%
Epithelioid hemangioma	0/5	0%	0/4	0%
Arteriovenous malformation	0/2	0%	0/2	0%
Angiomatosis	0/1	0%	0/1	0%
Hemangioma, NOS	0/3	0%	0/3	0%
Capillary/pyogenic hemangioma	0/5	0%	0/5	0%
Cavernous hemangioma	0/5	0%	0/5	0%
Juvenile hemangioma	0/1	0%	0/1	0%
Spindle cell hemangioma	0/4	0%	0/4	0%
Synovial hemangioma	0/1	0%	0/1	0%
Intramuscular hemangioma	0/6	0%	0/5	0%
Littoral cell hemangioma	0/6	0%	0/2	0%
Malignant hemangiopericytoma	0/1	0%	0/1	0%
Hemangiopericytoma, NOS	0/1	0%	0/1	0%
Sinonasal hemangiopericytoma	0/1	0%	0/1	0%
Glomus tumor	0/1	0%	0/1	0%
Atypical glomus tumor	0/2	0%	0/2	0%
Lymphangioma	0/7	0%	0/7	0%
Lymphangi leiomyomatosis	0/1	0%	0/1	0%
Papillary endothelial hyperplasia	0/2	0%	0/2	0%
Total cases	165		151	

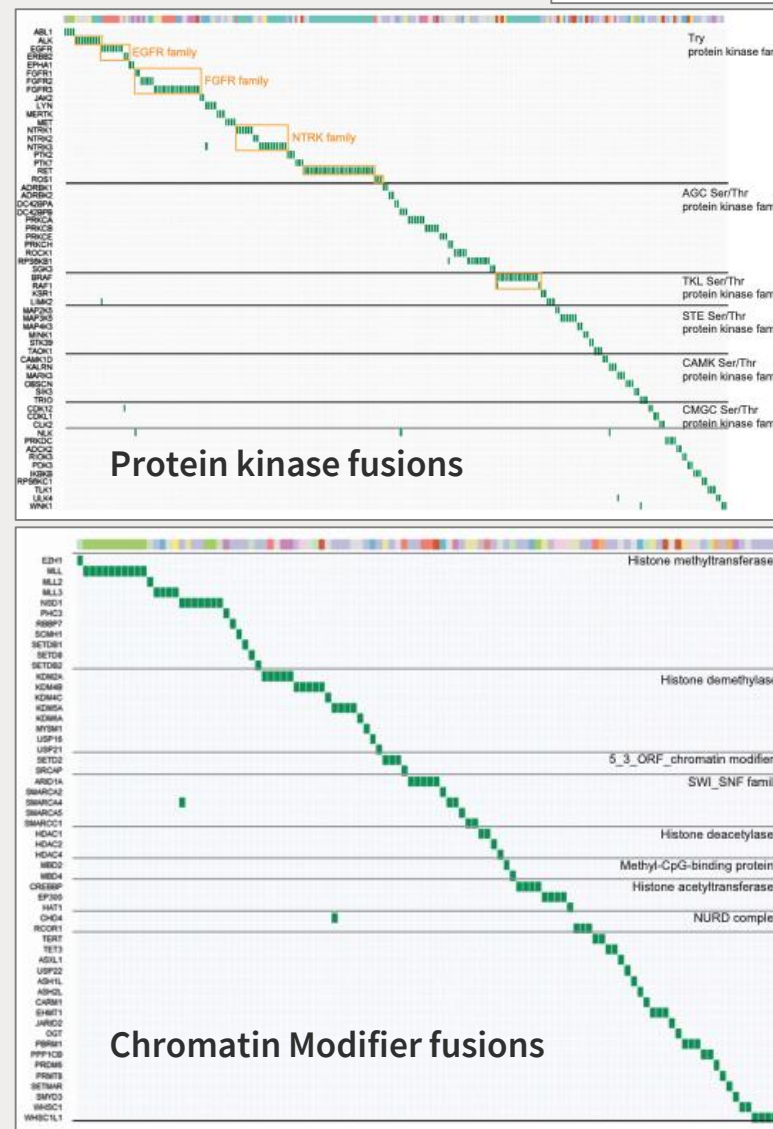
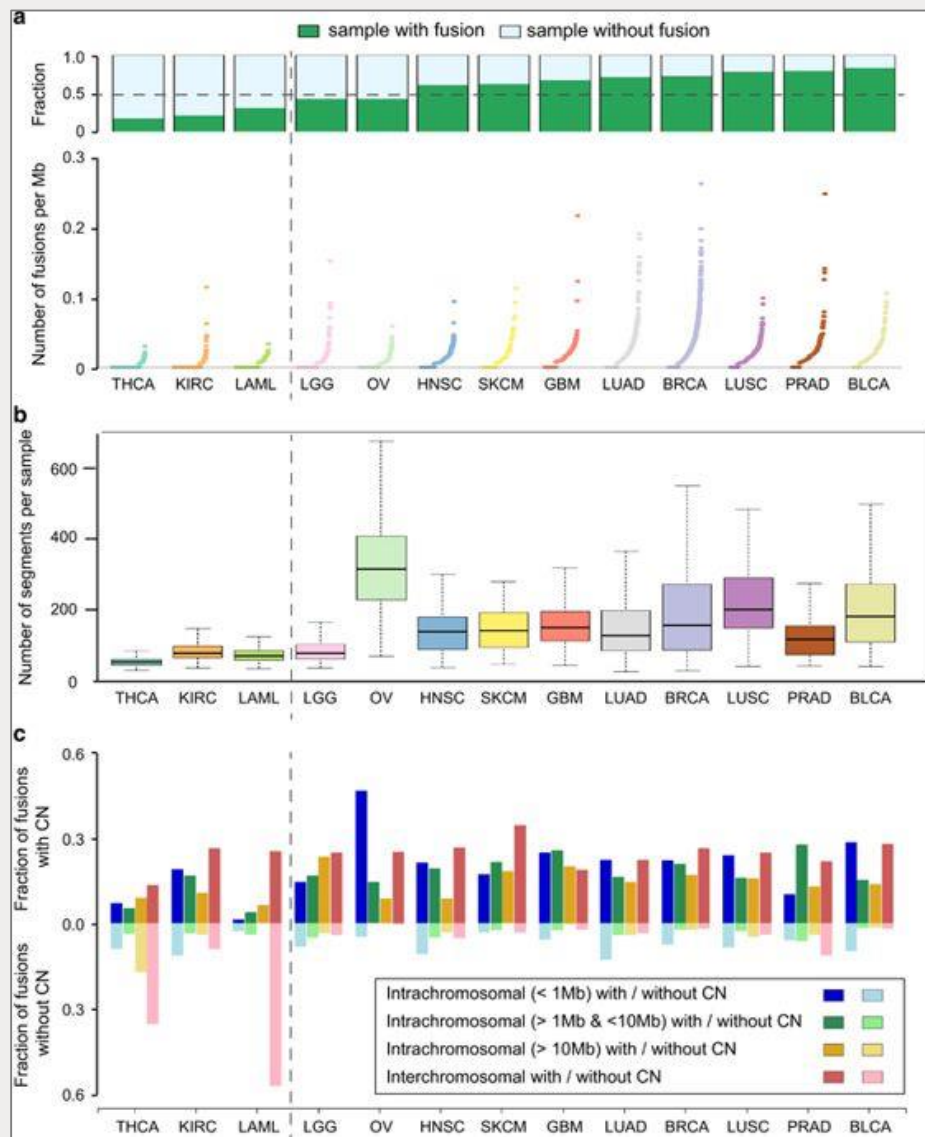
# Fusion supporting reads



# Spectrum of fusions in cancer types

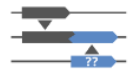
ORIGINAL ARTICLE  
 The landscape and therapeutic relevance of cancer-associated transcript fusions

K Yoshihara<sup>1,2</sup>, Q Wang<sup>1</sup>, W Torres-Garcia<sup>1</sup>, S Zheng<sup>1</sup>, R Vegesna<sup>1</sup>, H Kim<sup>1</sup> and RGW Verhaak<sup>1,3</sup>



# “Targeted” fusion detection methods

## ARCHER® FUSIONPlex® NGS Assays



### Novel fusions

AMP™ chemistry utilizes open-ended targeted amplification to identify gene fusions whether or not the fusion partner is known [Video](#) [Blog](#)



### Expression

Molecular barcodes coupled with open-ended amplification allows for determination of RNA vs DNA reads [Blog](#)

## nanoString

- Breast Cancer 360™ Panel
- PanCancer IO 360™ Panel
- CAR-T Characterization Panel
- Hallmarks of Cancer Collection
  - PanCancer Pathways Panel
  - PanCancer Immune Profiling Panel
  - PanCancer Progression Panel
- PlexSet™ Pre-selected Panels
- Vantage 3D™ RNA Panels
- Vantage 3D™ Gene Fusion Panels
- miRNA Panels
- Kinase Panel
- Stem Cell Panel
- nCounter Gene Fusion Panels (Ex-US only)

Prepare Library | Sequence | Analyze Data **illumina®**

## TruSight™ Oncology 500

	NTRK1, NTRK2, NTRK3 (pan-cancer)   MSI (pan-cancer)								
	Lung	Melanoma	Colon	Ovarian	Breast	Gastric	Bladder	Myeloid	Sarcoma
	AKT1 ALK BRAF DDR2 EGFR ERBB2 FGFR1 FGFR3 KRAS MAP2K1 MET NRAS PIK3CA PTEN RET TP53 TMB	BRAF CTNNB1 GNA11 GNAQ KIT MAP2K1 NF1 NRAS PDGFRA PIK3CA PTEN TP53	AKT1 BRAF HRAS KRAS KIT MLH1 MSH2 MSH6 NRAS PIK3CA PTEN SMAD4 TP53	BRAF BRCA1 BRCA2 KRAS PDGFRA FOXL2 TP53	AKT1 AR BRCA1 BRCA2 ERBB2 FGFR1 FGFR2 PIK3CA PTEN	BRAF KIT KRAS MET MLH1 PDGFRA TP53	MSH6 PMS2 TSC1	ABL1 ASXL1 CALR CEBPA ETV6 EZH2 FLT3 GATA2 IDH1 IDH2 JAK2 KIT MPL NPM1 RUNX1 SF3B1 SRSF2 TP53	ALK APC BRAF CDK4 CTNNB1 ETV6 EWSR1 FOXO1 GLI1 KIT MDM2 MYOD1 NAB2 NF1 PAX3 PAX7 PDGFRA PDGFRB SDHB SDHC SMARCB1 TFE3 WT1



# FDA-approved drugs targeting oncogenic fusions in solid tumors

Fusion target	Therapy	Indication	FDA approval
ALK fusion	Crizotinib	Lung	August 2011
	Ceritinib	Lung	May 2017
	Alectinib	Lung	November 2017
	Brigatinib	Lung	May 2020
	Lorlatinib	Lung (second line)	November 2018
FGFR fusion	Erdafitinib	Urothelial	April 2019
	Pemigatinib	Cholangiocarcinoma	April 2020
ROS1 fusion	Crizotinib	Lung	March 2016
	Entrectinib	Lung	August 2019
RET fusion	Selpercatinib	Lung Thyroid	May 2020
	Pralsetinib	Lung	September 2020
NTRK1/2/3 fusion	Larotrectinib	Solid tumor	November 2018
	Entrectinib	Solid tumor	August 2019
PDGFB fusion	Imatinib	DFSP	November 2006
MET exon 14 skipping	Capmatinib	Lung	May 2020

# Tools for detecting fusion transcripts

## Gene fusion detection software tools | RNA sequencing

High-throughput sequencing software tools > RNA sequencing software tools

<http://omictools.com/gene-fusion-detection-category>

### RNA-seq short-reads “only”

Bellerophon  
BreakFusion  
chimeraScan  
CRAC  
deFuse  
EricScript  
FusionAnalyser  
FusionCatcher  
FusionFinder  
FusionHunter  
FusionQ  
FusionSeq  
Jaffa  
MapSplice  
PRADA  
shortFuse  
SnowShoes-FTD  
SOAPFuse/Fusion  
TopHat-Fusion  
STAR-fusion

### RNA-seq & DNA-seq

BreakTrans  
Comrad  
nFuse

### Gene fusion annotation

Chimera  
Pegasus

## De novo transcriptome assembly software tools | RNA sequencing

High-throughput sequencing software tools > RNA sequencing software tools

<http://omictools.com/transcriptome-assembly-category>

### Transcript Assembly

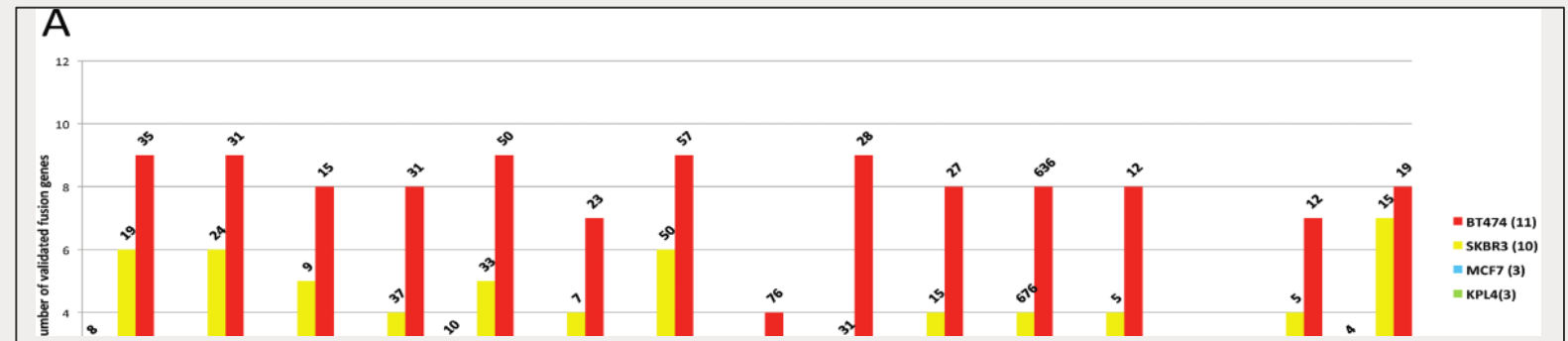
CuffLinks  
Scripture  
Trinity  
Trans-Abyss

Published online 17 November 2015

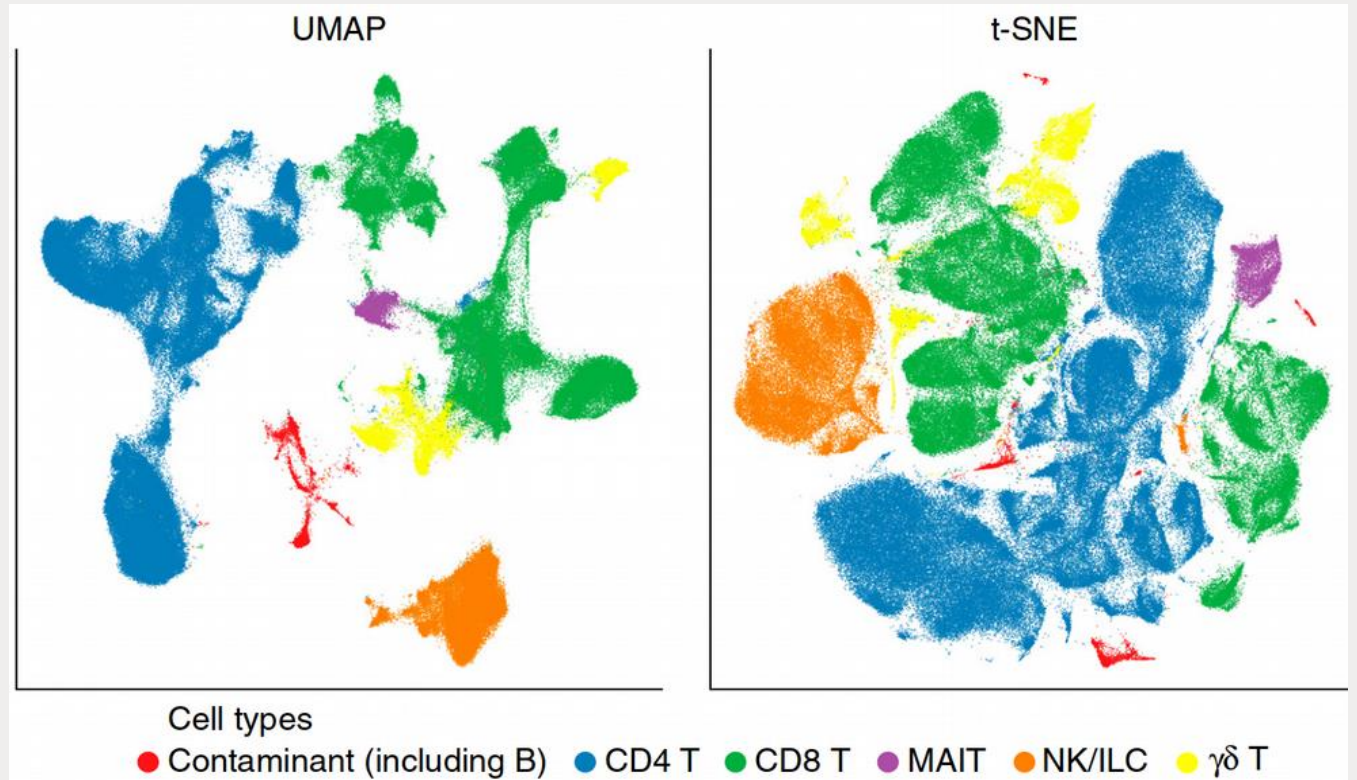
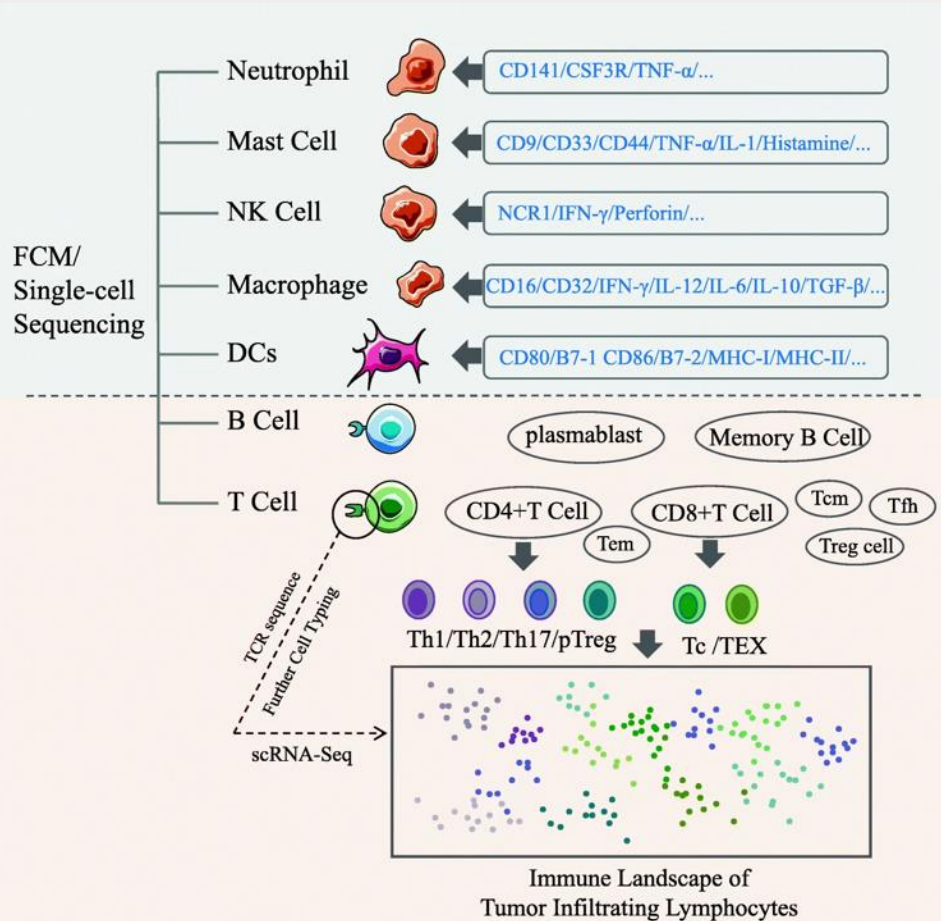
*Nucleic Acids Research*, 2016, Vol. 44, No. 5 e47  
doi: 10.1093/nar/gkv1234

## Comprehensive evaluation of fusion transcript detection algorithms and a meta-caller to combine top performing methods in paired-end RNA-seq data

Silvia Liu<sup>1,2,†</sup>, Wei-Hsiang Tsai<sup>3,†</sup>, Ying Ding<sup>1,2,†</sup>, Rui Chen<sup>1</sup>, Zhou Fang<sup>1</sup>, Zhiguang Huo<sup>1</sup>, SungHwan Kim<sup>1</sup>, Tianzhou Ma<sup>1</sup>, Ting-Yu Chang<sup>4</sup>, Nolan Michael Priedigkeit<sup>5</sup>, Adrian V. Lee<sup>6</sup>, Jianhua Luo<sup>7</sup>, Hsei-Wei Wang<sup>3,4,8,\*</sup>, I-Fang Chung<sup>3,8,\*</sup> and George C. Tseng<sup>1,2,\*</sup>



# Single-cell RNA sequencing (scRNA-seq): higher magnification using the NGS microscope

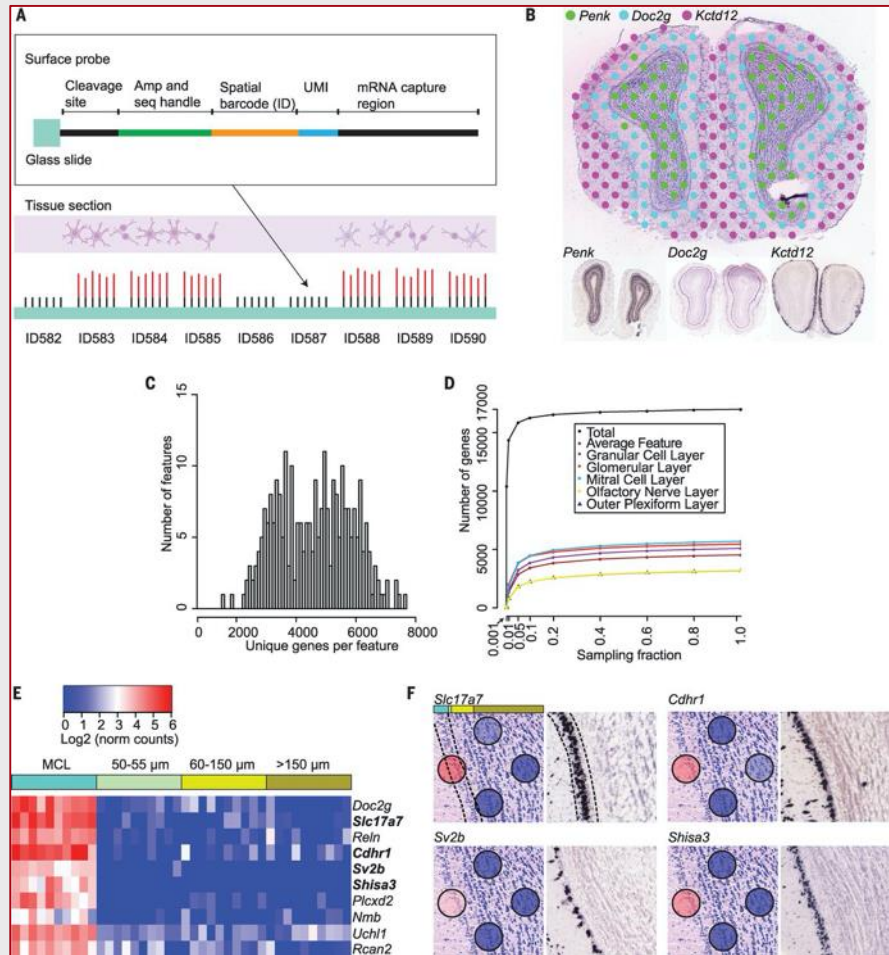


<https://towardsdatascience.com/reduce-dimensions-for-single-cell-4224778a2d67>

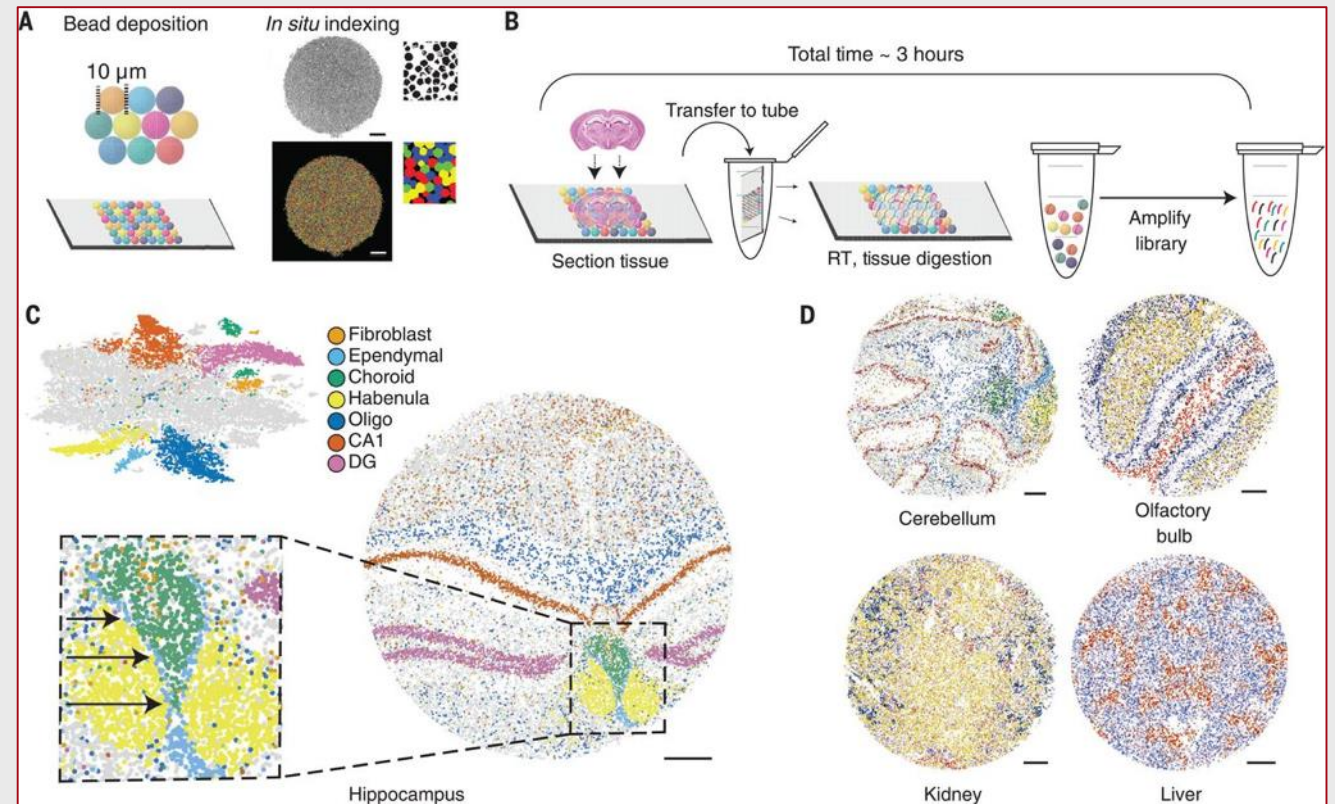
Zhang et al. *Journal of Experimental & Clinical Cancer Research* (2021) 40:81  
<https://doi.org/10.1186/s13046-021-01874-1>

# Spatial profiling

Measurements (such as gene expression) that maintain the spatial information.



P. L. Ståhl et al., *Science*. 353, 78–82 (2016).




S. G. Rodrigues et al., *Science*. 363, 1463–1467 (2019).

# Summary and future directions

- Massively Parallel Sequencing has enabled the discovery of additional fusion transcripts
  - Specificity is the main challenge: too many false positives (FPs)!
  - Longer reads: could help overcome the limitations of short reads
  - Combination of tools may help further improve on the reduction of FPs
- “For the large bioinformatics community, development of a high-performing (accurate and fast) fusion detection tool or methods to combine top-performing tools remains an important and open question”

✉ [ans2077@med.cornell.edu](mailto:ans2077@med.cornell.edu)

🐦 @asbonerw

 **Weill Cornell Medicine**  
Caryl and Israel Englander  
Institute for Precision Medicine

 **Weill Cornell Medicine**  
Pathology & Laboratory Medicine

 **Weill Cornell Medicine**  
Clinical Genomics Lab

 **New York Presbyterian**